

III Jornadas de genómica clínica  
FCEN-UBA  
25 de octubre de 2024

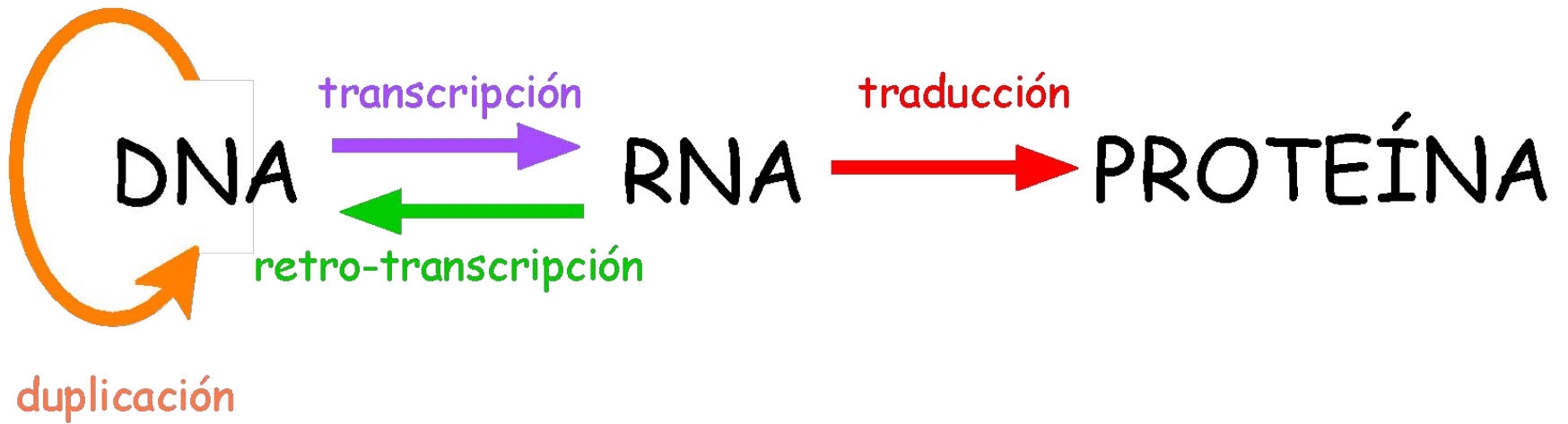
*Terapia combinada de una enfermedad hereditaria sobre la base  
de modulación del splicing alternativo y del estado de la cromatina*

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Universidad de Buenos Aires - Argentina



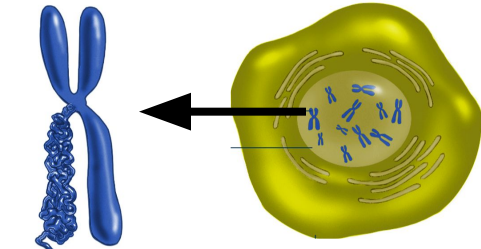
Los principales productos de los genes son proteínas

¿Cómo se fabrican?



cromosoma

célula



gen

ADN

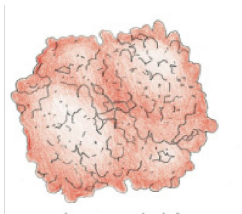


TRANSCRIPCIÓN



Ácido ribonucleico (ARN)

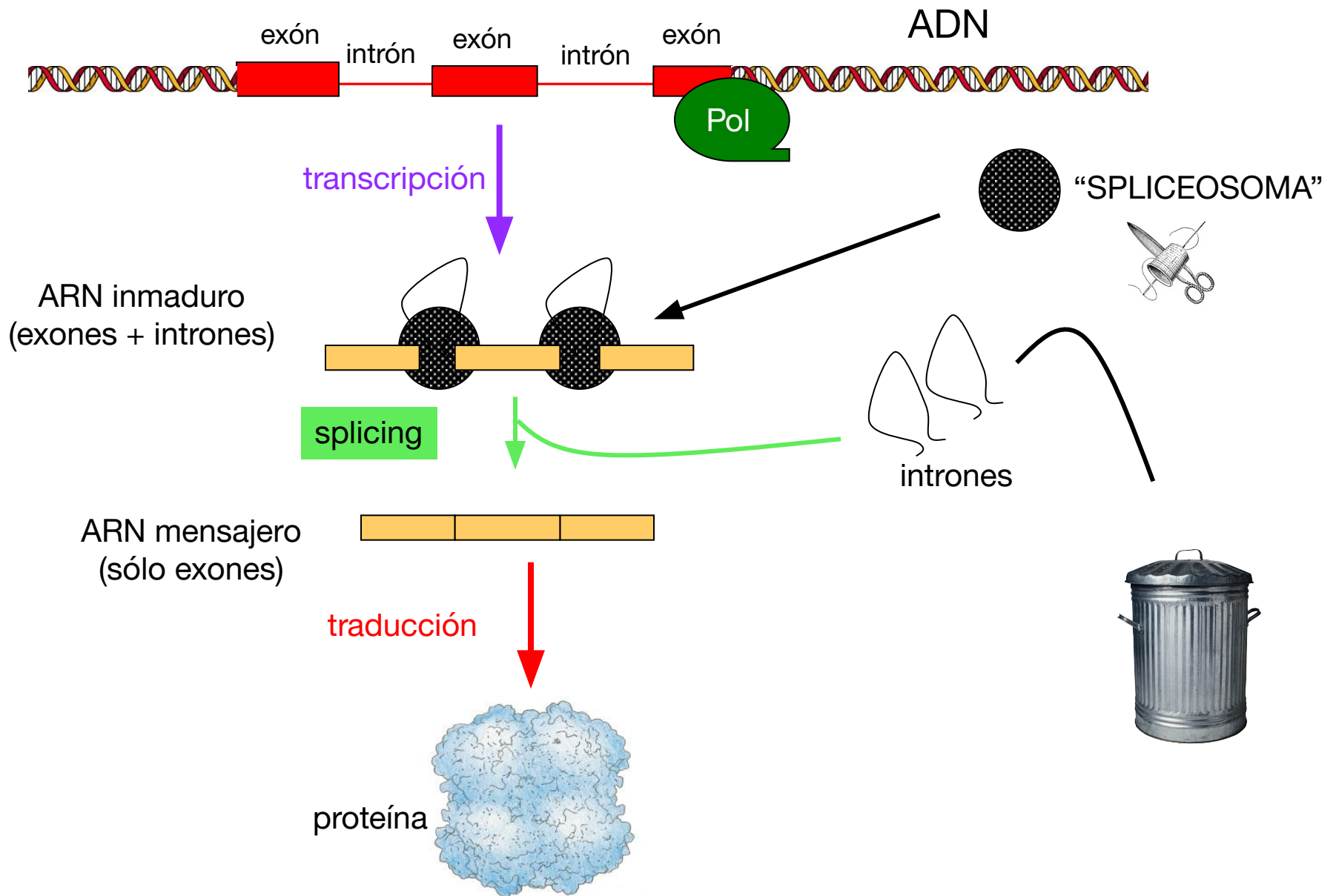
TRADUCCIÓN



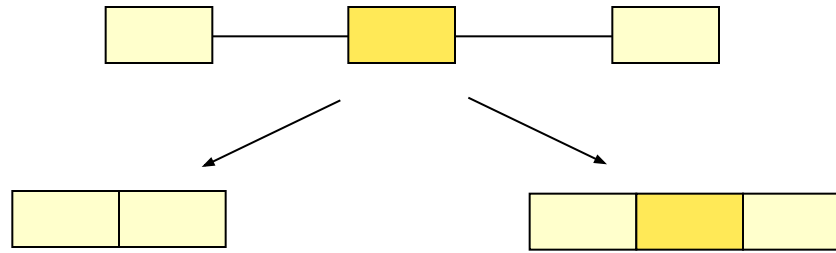
proteína

función

# Un gen



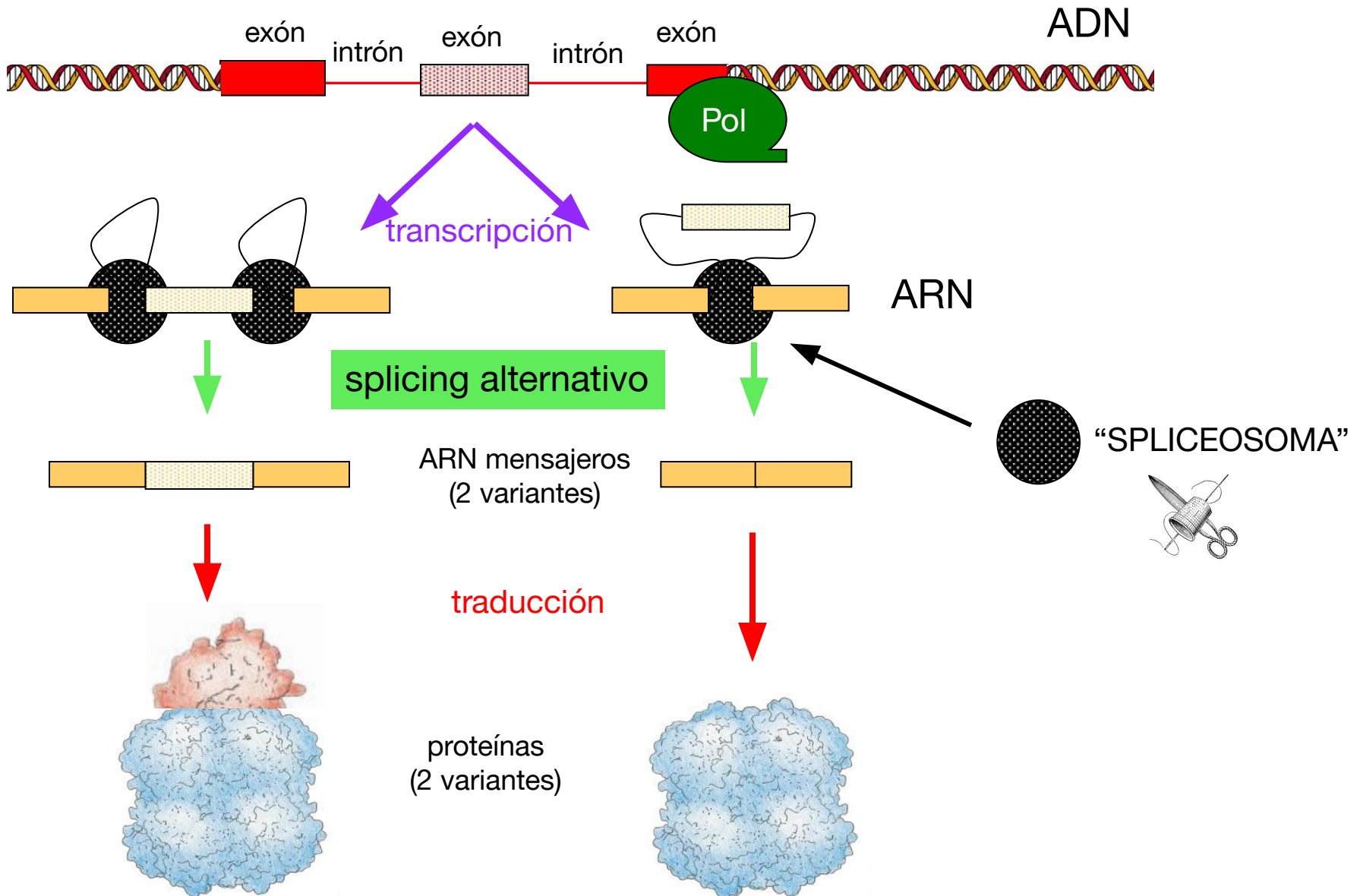
*Splicing alternativo*

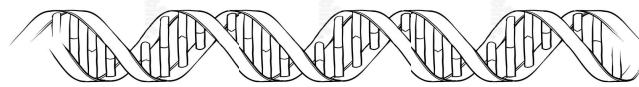


## *Alternative splicing*

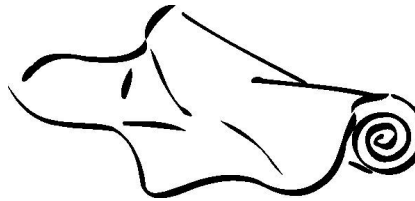
- 1. It is more a rule than an exception. It is estimated to affect the expression of nearly 95% of human genes.*
- 2. It explains how a vast protein diversity is achieved with a limited number of genes.*
- 3. Mutations that affect alternative splicing regulatory sequences (splicing enhancers and silencers) are a widespread source of human disease.*
- 4. Alternative splicing regulation not only depends on the interaction of splicing factors with their target sequences in the pre-mRNA but is coupled to RNA polymerase II (pol II) transcription.*

# Un gen

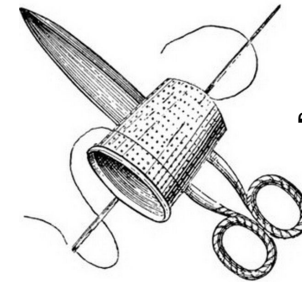




ADN (gen)

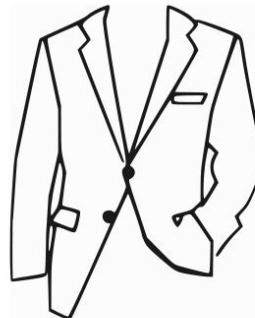


ARN mensajero precursor



“SPLICEOSOMA”

NÚCLEO



Single-breasted



Double-breasted

2 ARN mensajeros diferentes

CITOPLASMA



2 proteínas diferentes

un gen → una proteína

un gen → muchas proteínas

El splicing alternativo parece ser la causa de la gran **complejidad** de los vertebrados (nosotros)

# Gusano *Caenorhabditis elegans*

Invertebrado microscópico de 1 mm de largo formado por 1000 células



**19.000** genes en cada célula

Fuente: <http://www.bio.unc.edu/faculty/goldstein/lab/movies.html>

## *Homo sapiens sapiens*

Vertebrado macroscópico de casi 2 m de largo formado por  $10^{13}$  células

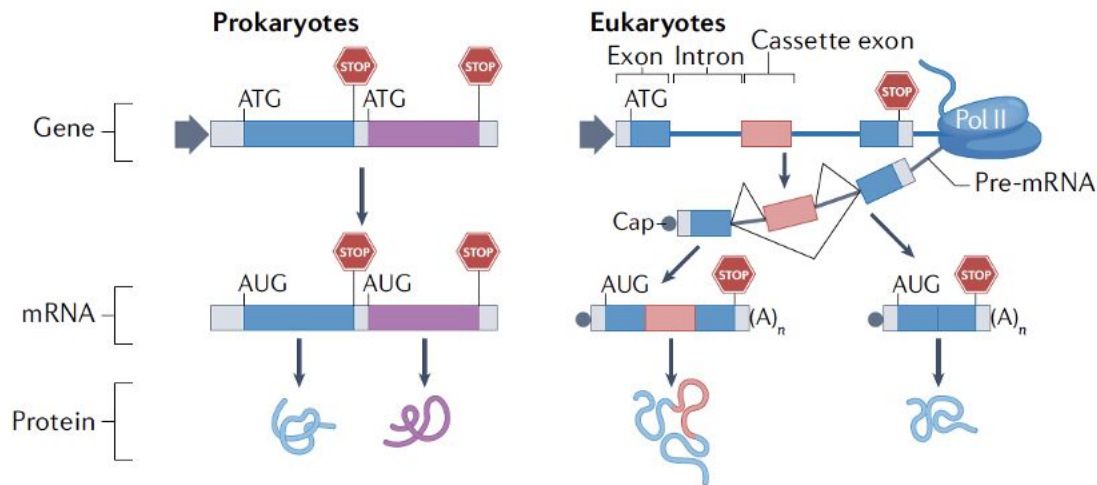


**21.000** genes en cada célula

# The physiology of alternative splicing

Luciano E. Marasco<sup>1,2</sup> and Alberto R. Kornblihtt<sup>1</sup>  

NATURE REVIEWS | **MOLECULAR CELL BIOLOGY** 24, 242–254 (2023)

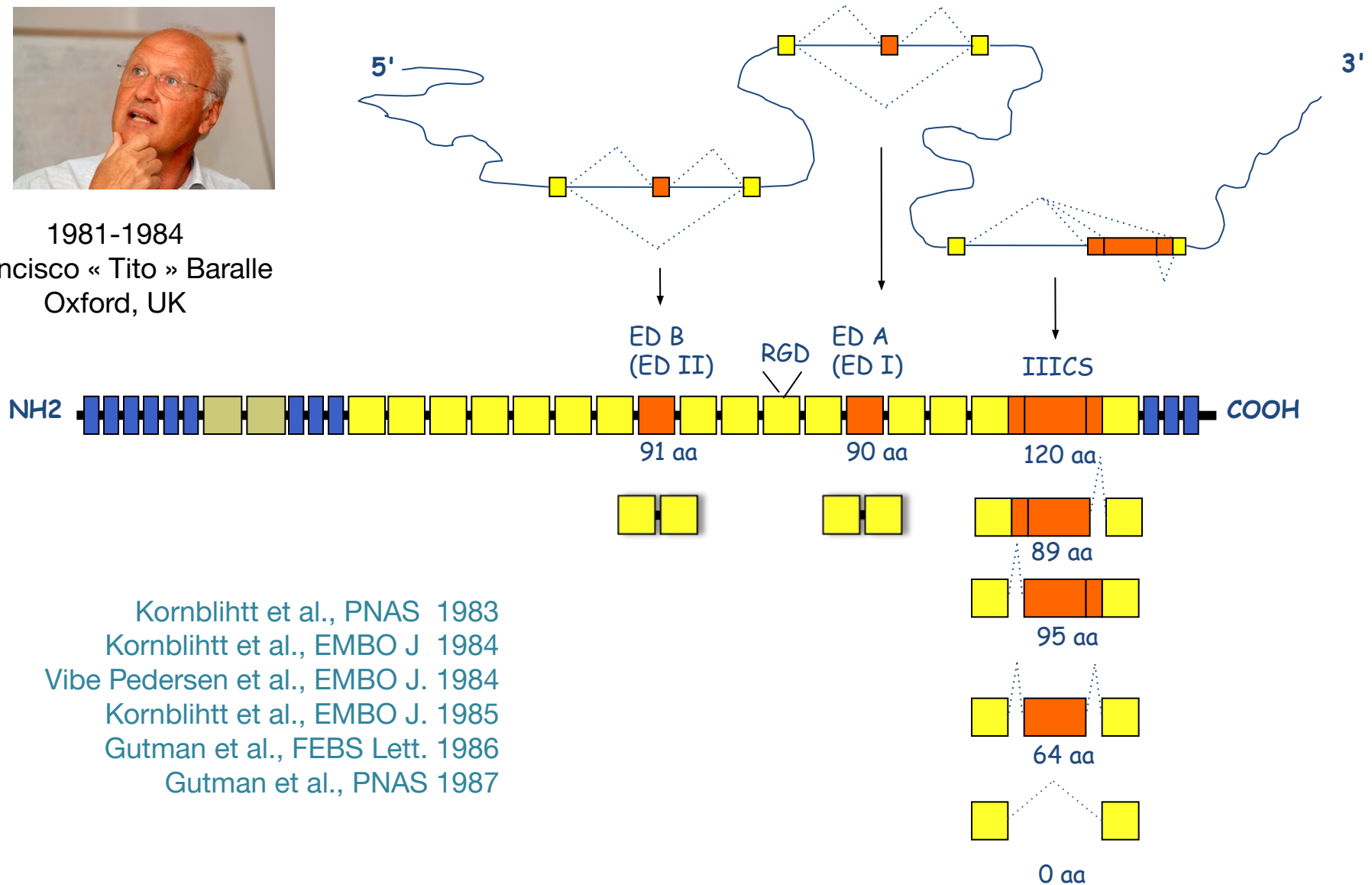


# Fibronectina humana



1981-1984

Francisco « Tito » Baralle  
Oxford, UK



Kornblihtt et al., PNAS 1983

Kornblihtt et al., EMBO J 1984

Vibe Pedersen et al., EMBO J. 1984

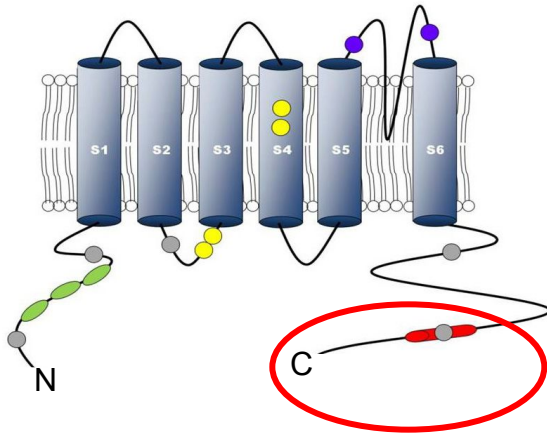
Kornblihtt et al., EMBO J. 1985

Gutman et al., FEBS Lett. 1986

Gutman et al., PNAS 1987



## TRPV1 (CANAL IÓNICO DE NEURONAS)

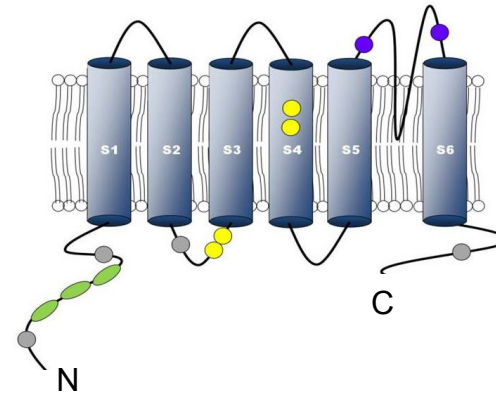


Ganglios de la raíz dorsal

>43 °C

CALOR NOCIVO

DOLOR



Ganglios del trigémino

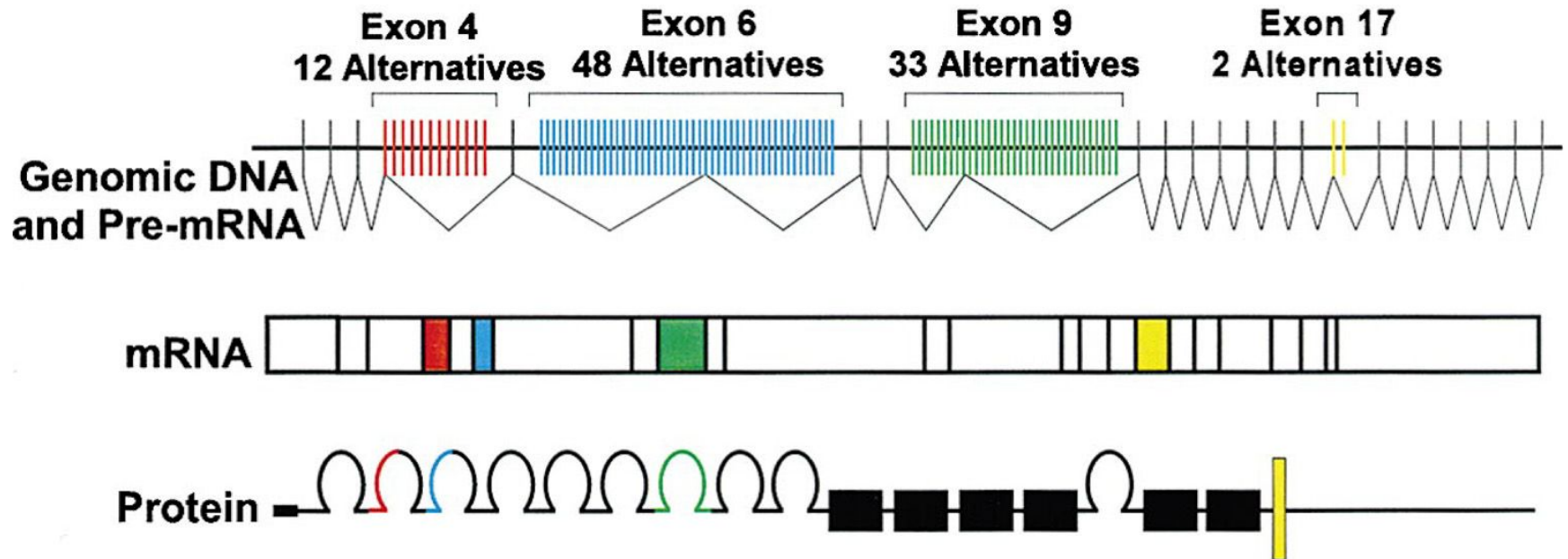
>30 °C

SENSADO DE INFRARROJO

ALIMENTACIÓN

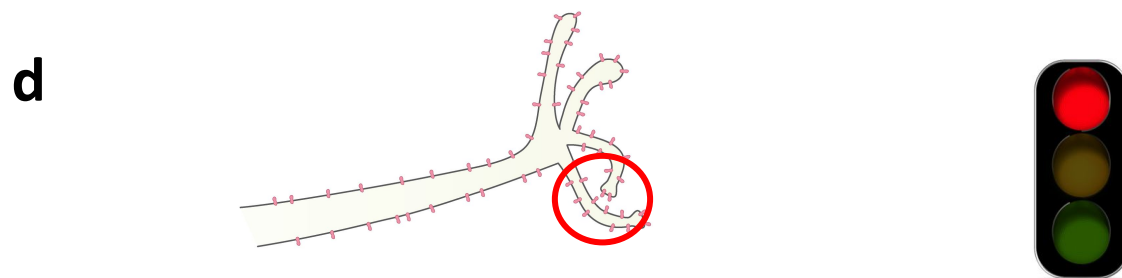
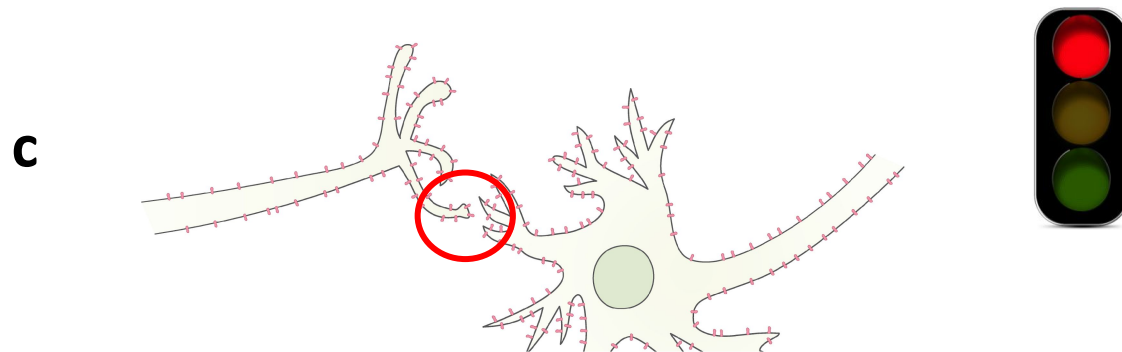
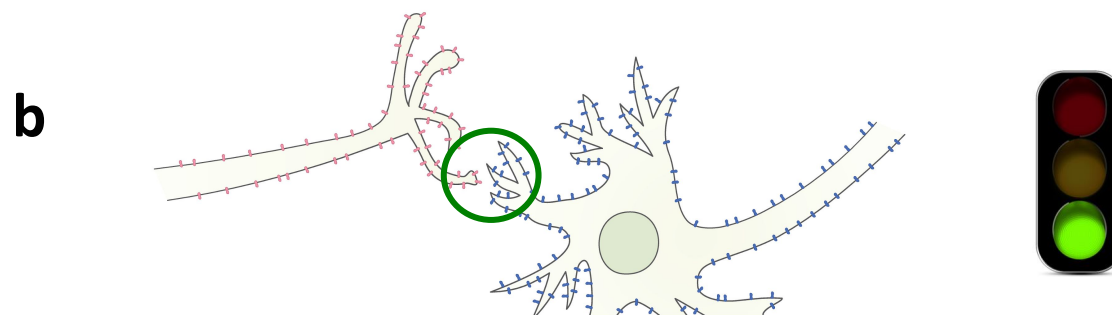
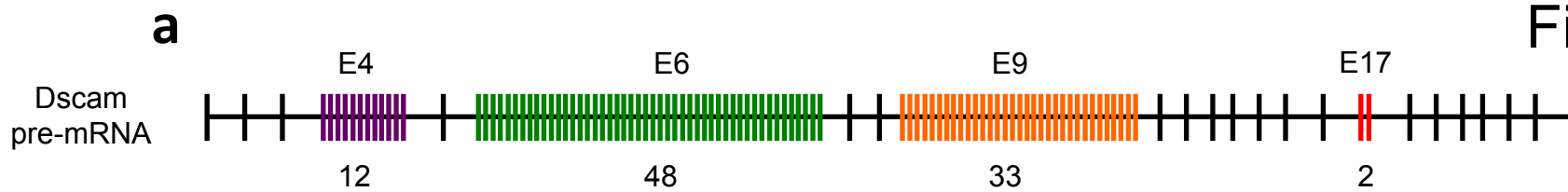
# *Gene de la mosca DSCAM*

## *38.016 variantes de mRNA*

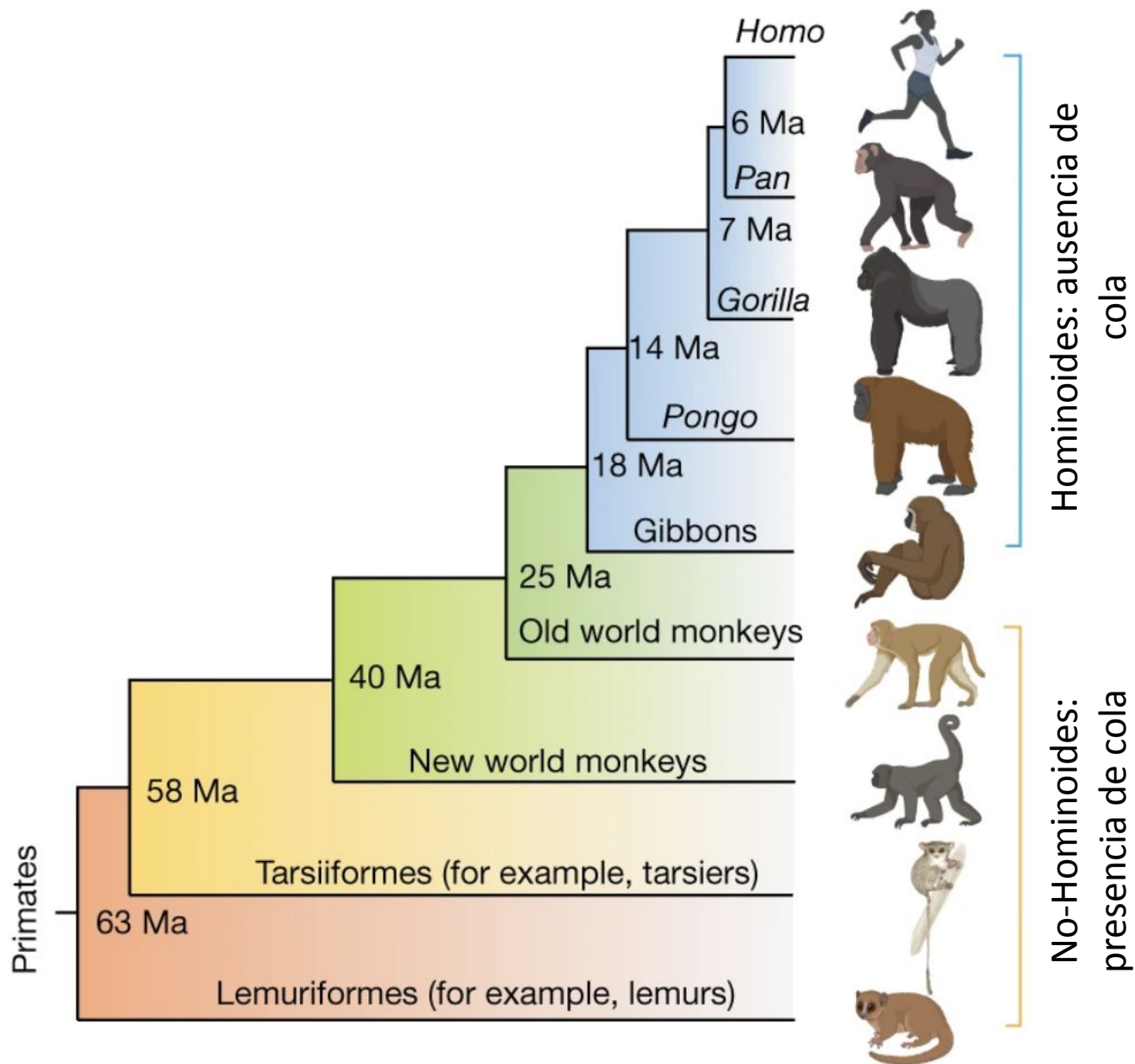


Schmucker et al. *Cell* 101, 671-684 (2000)

Fig. 3



El splicing alternativo es la causa de la pérdida de la cola en los primates hominoides (gibones, orangutanes, gorilas, chimpancés y humanos).  
Nature, febrero 2024.



[nature](#) > [articles](#) > article

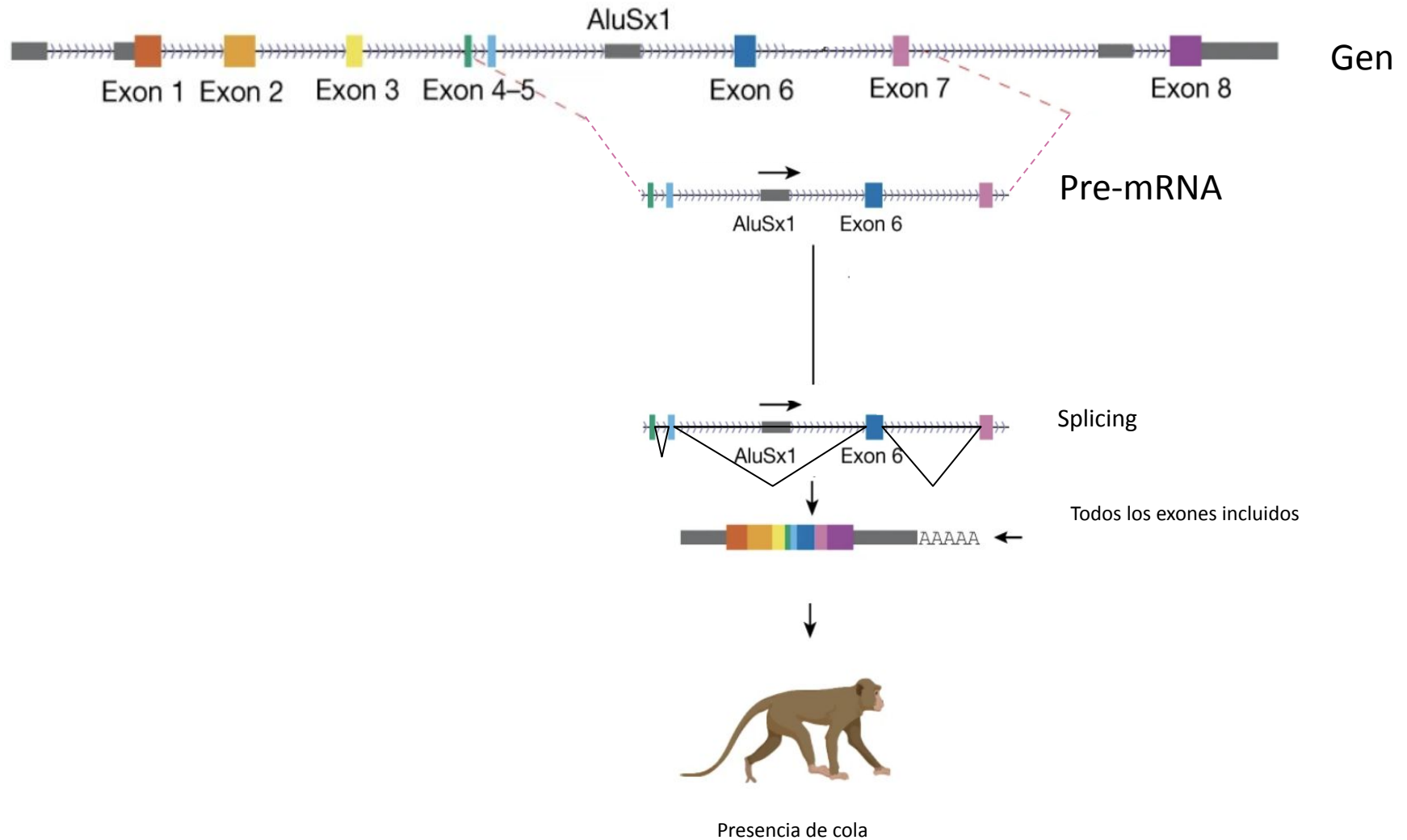
Article | [Open access](#) | Published: 28 February 2024

## On the genetic basis of tail-loss evolution in humans and apes

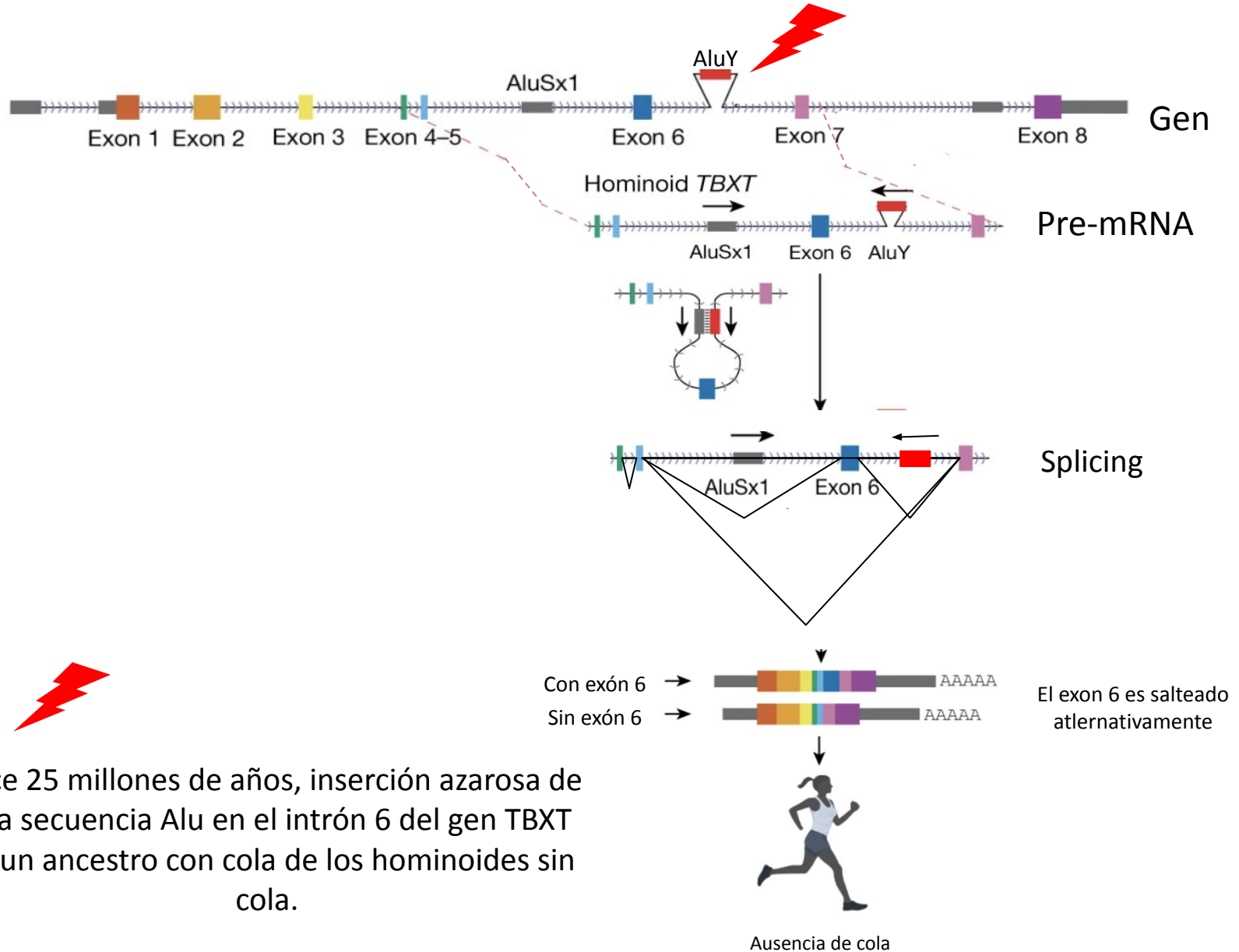
[Bo Xia](#) , [Weimin Zhang](#), [Guisheng Zhao](#), [Xinru Zhang](#), [Jiangshan Bai](#), [Ran Brosh](#), [Aleksandra Wudzinska](#), [Emily Huang](#), [Hannah Ashe](#), [Gwen Ellis](#), [Maayan Pour](#), [Yu Zhao](#), [Camila Coelho](#), [Yinan Zhu](#), [Alexander Miller](#), [Jeremy S. Dasen](#), [Matthew T. Maurano](#), [Sang Y. Kim](#), [Jef D. Boeke](#)  & [Itai Yanai](#) 

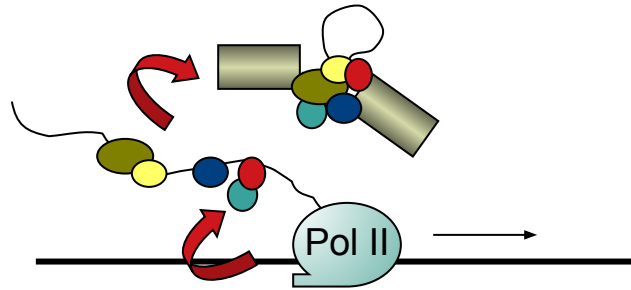
[Nature](#) **626**, 1042–1048 (2024) | [Cite this article](#)

## Gen TBXT en primates no hominoides



# Gen TBXT en primates hominoides





*Splicing and alternative splicing are coupled to  
RNA polymerase II transcription*

Cramer et al. PNAS 1997  
Cramer et al. Molecular Cell 1999

*Modes of coupling*

*changes in pol II elongation rate  
(kinetic coupling)*

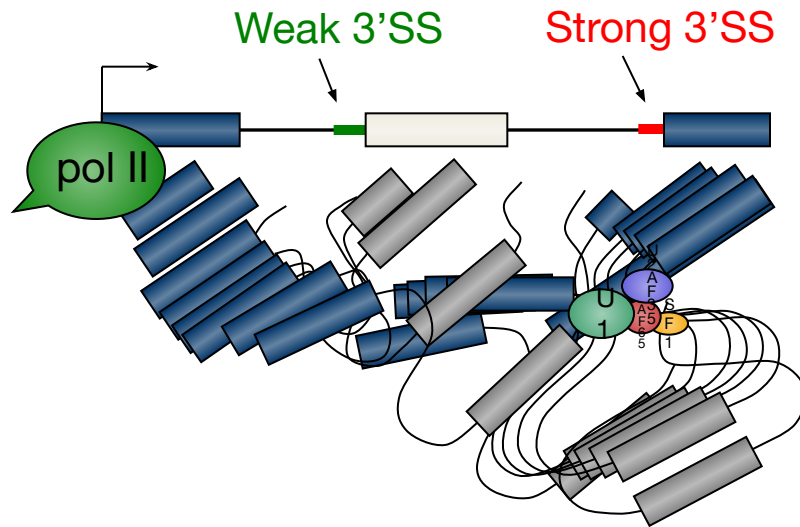
*and/or*

*recruitment of processing factors  
to RNA polymerase II (CTD),  
chromatin or nascent RNA  
(recruitment coupling)*

*Slow elongation, higher exon  
inclusion*

# First come, first served

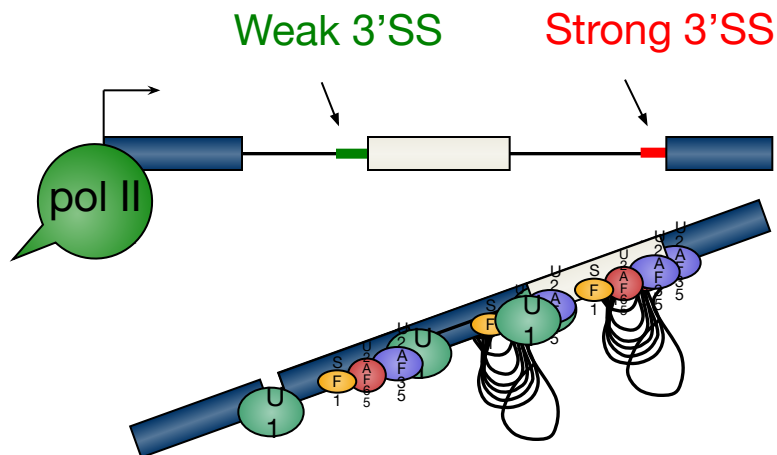
*Fast elongation/no pauses*



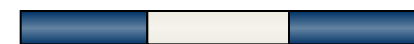
Exclusion



*Slow elongation/ with pauses*



Inclusion



Kadener *et al.* EMBO J. 2001  
Nogués *et al.* JBC 2002  
de la Mata *et al.* Mol. Cell 2003  
Fededa *et al.* Mol. Cell 2005  
Alló *et al.* NSMB 2009  
Muñoz *et al.* Cell 2009  
de la Mata *et al.* RNA 2010  
Dujardin *et al.* Mol. Cell 2014

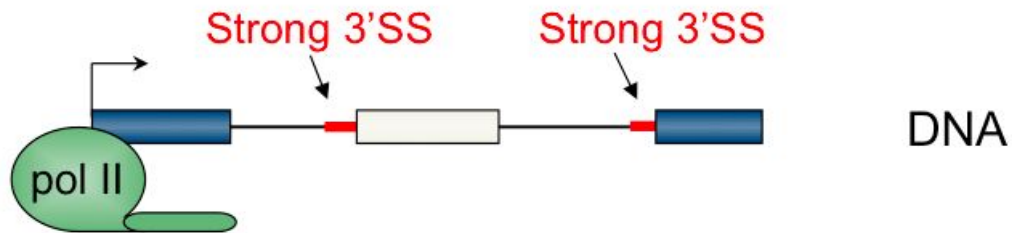
*Slow elongation, higher exon  
inclusion*

*50-80% of elongation-sensitive  
alternative splicing events*

*Slow elongation, higher exon  
skipping*

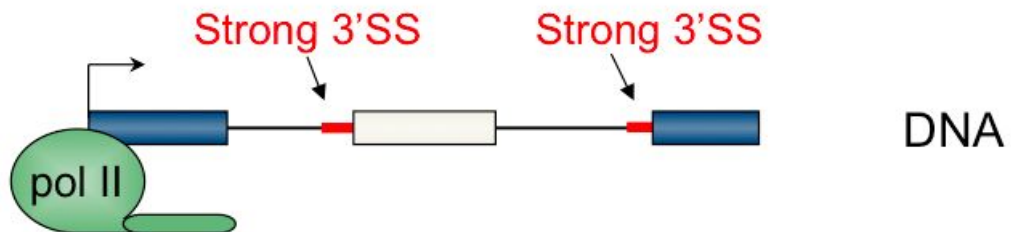
*20-50% of elongation-sensitive  
alternative splicing events*

## ***Fast elongation***

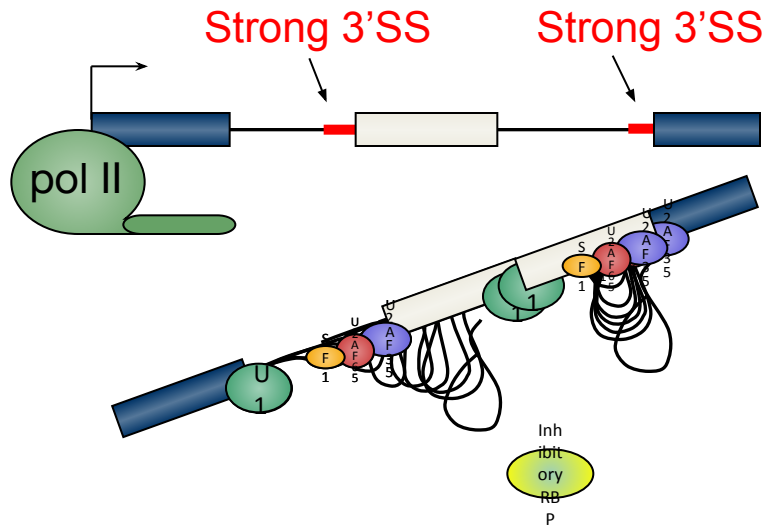


ETR-3

## ***Slow elongation/pauses***



## ***Fast elongation***



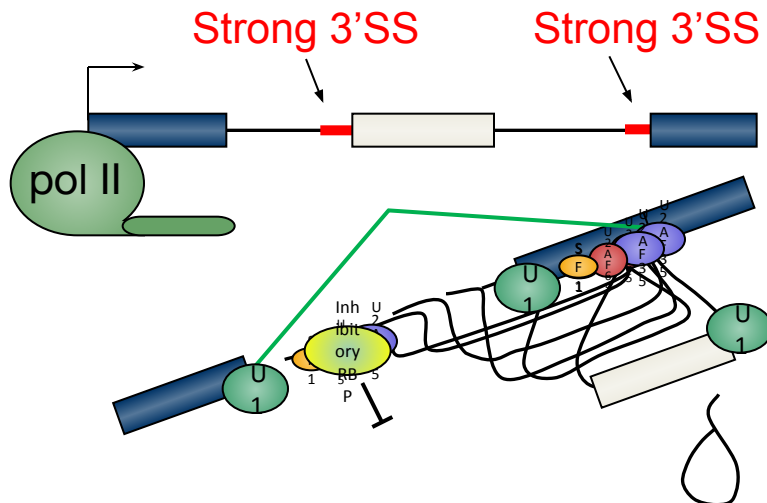
DNA



Inclusion



## ***Slow elongation/pauses***



DNA

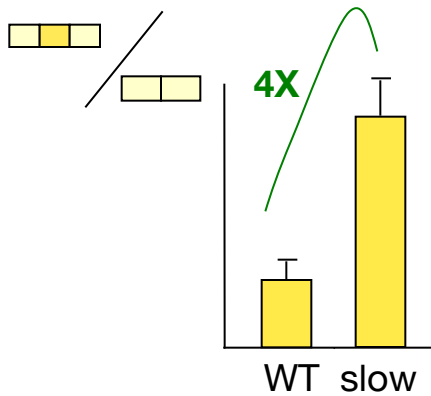
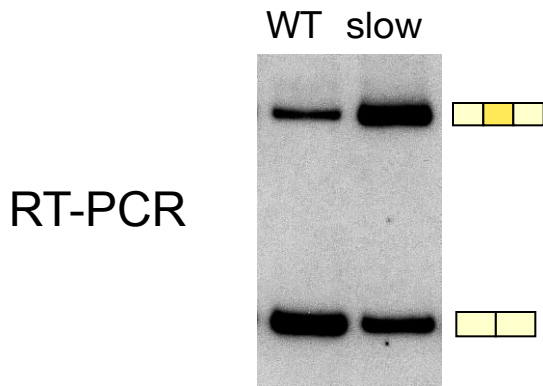


Skipping



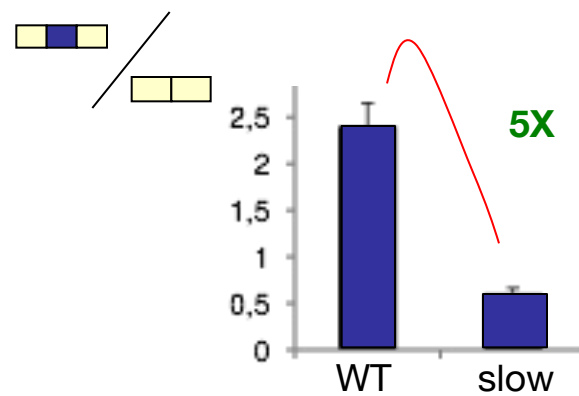
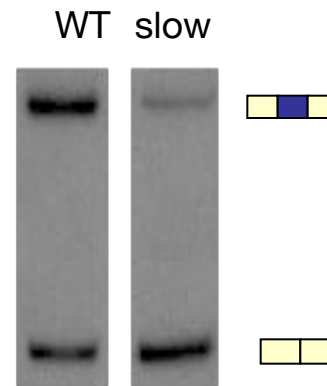
# Slow Pol II

## *FN E33 (EDI)*



de la Mata et al. Molecular Cell 2009

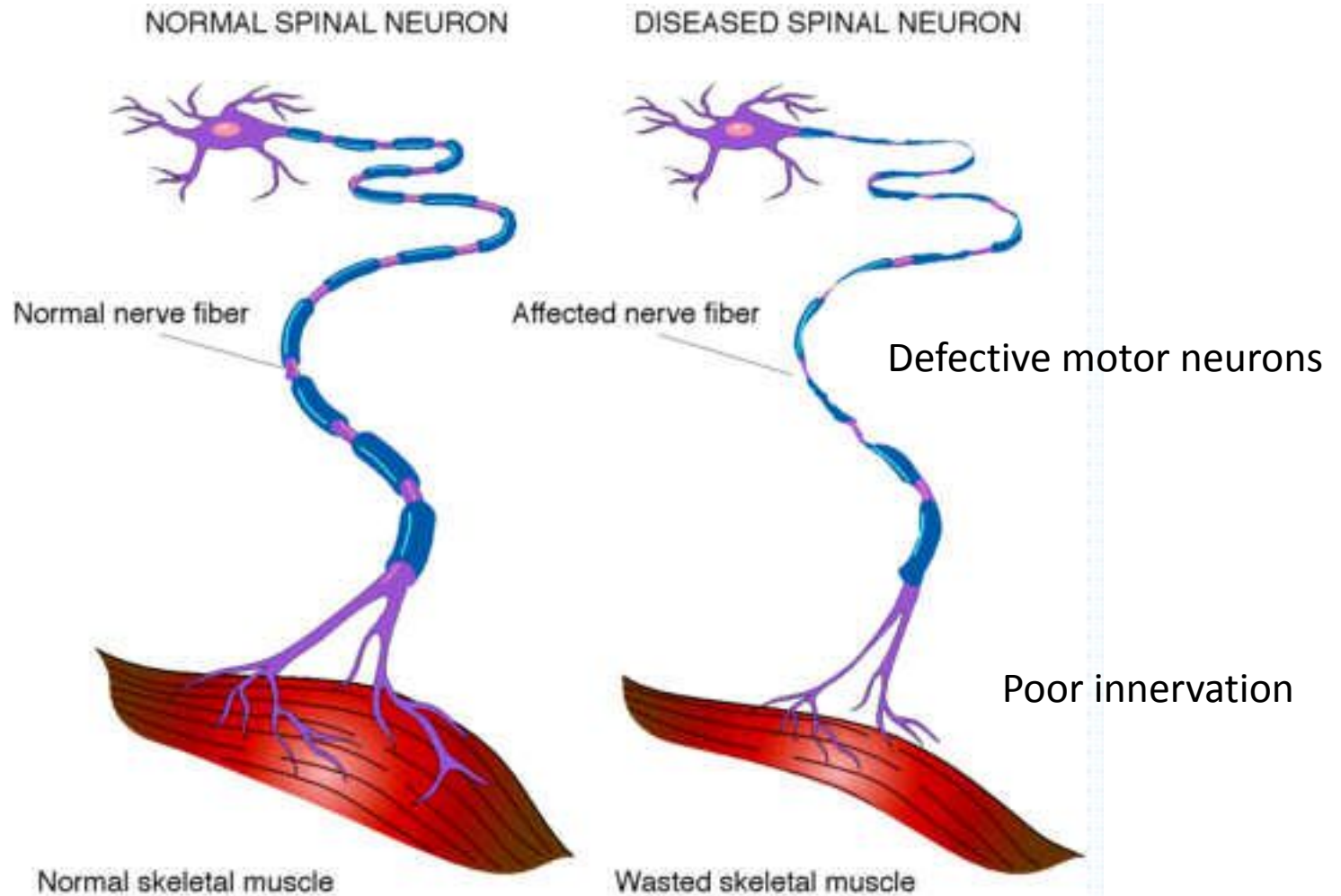
## *CFTR E9*



Dujardin et al. Molecular Cell 2014

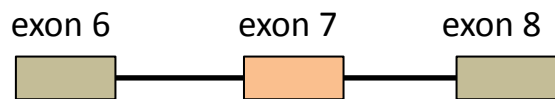
*SMA = Spinal muscular atrophy*

# SPINAL MUSCULAR ATROPHY (SMA) (autosomal recessive)



# SMN1 gene

DNA



transcription

Pre-mRNA



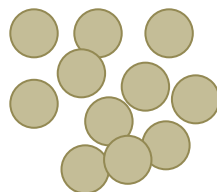
splicing

mRNA



translation

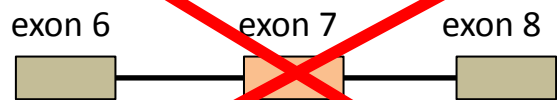
SMN protein  
(involved in  
splicing)



100%

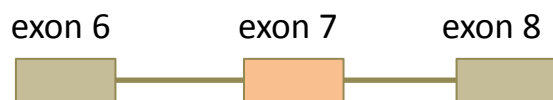
## SMN1 gene

DNA



transcription

Pre-mRNA



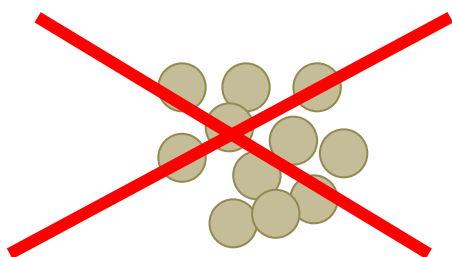
splicing

mRNA



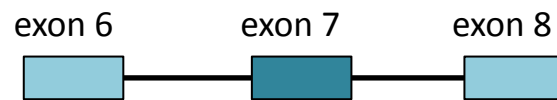
translation

SMN protein



100%

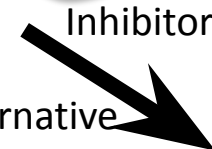
## SMN2 gene



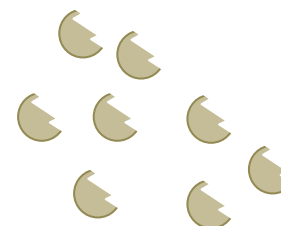
transcription



Alternative  
splicing



translation



20%

80%



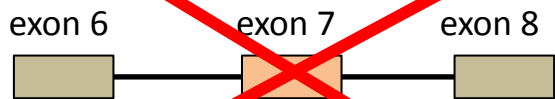
Adrian Krainer  
CSHL, New York

Spinraza (nusinersen) is an antisense oligonucleotide (ASO)

2'O (2-methoxyethyl) phosphorothyonate antisense oligonucleotide

## SMN1 gene

DNA



transcription

Pre-mRNA

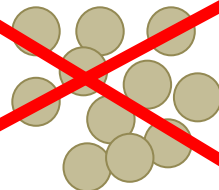


splicing

mRNA

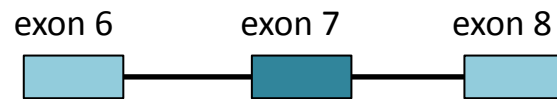


translation



100%

## SMN2 gene



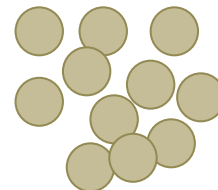
transcription



Nusinersen  
(Spinraza)



Alternative  
splicing



80%



20%



Adrian Krainer  
CSHL, New York

3 years old riding a tricycle  
1 year old, holding a heavier toy

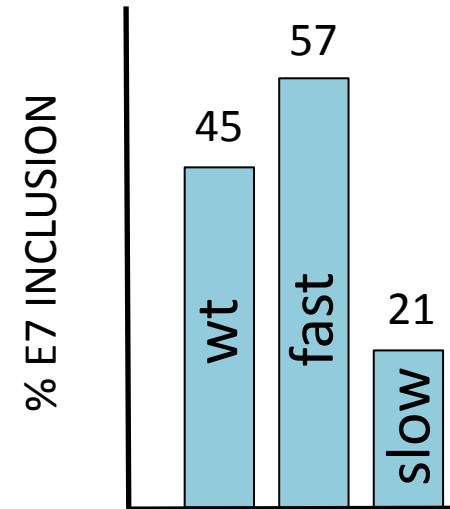
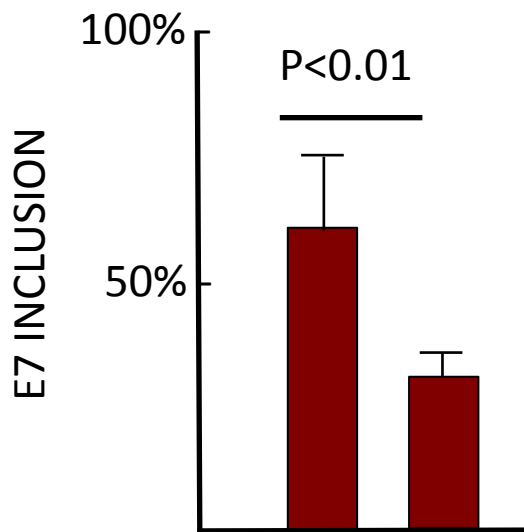
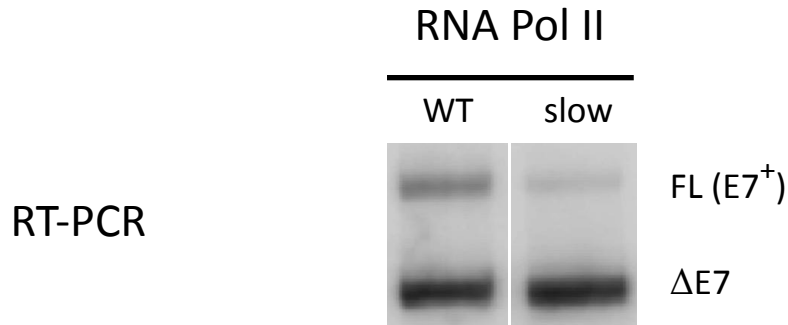
*Is SMN2 E7 alternative splicing  
controlled by Pol II elongation?*

Spinraza = nusinersen  
ccagcattattgaaagtga

ASO1  
ccagcattattgaaagtgaat

# *SMN2 exon 7 is skipped when elongation is slow*

HEK293 cells



Bentley lab: Fong et al. 2014

*SMN2 exon 7 is a  
class II exon*

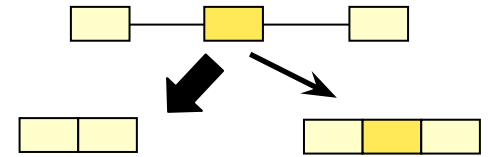
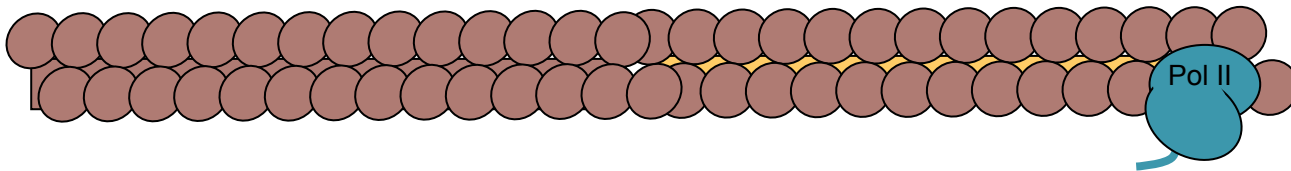
Marasco et al., Cell 2022

*SMN2 E7 is a type II exon*

*Slow elongation, higher exon  
skipping*

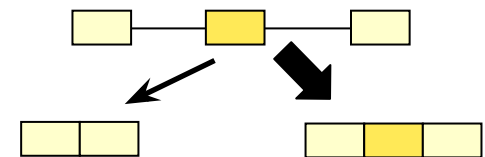
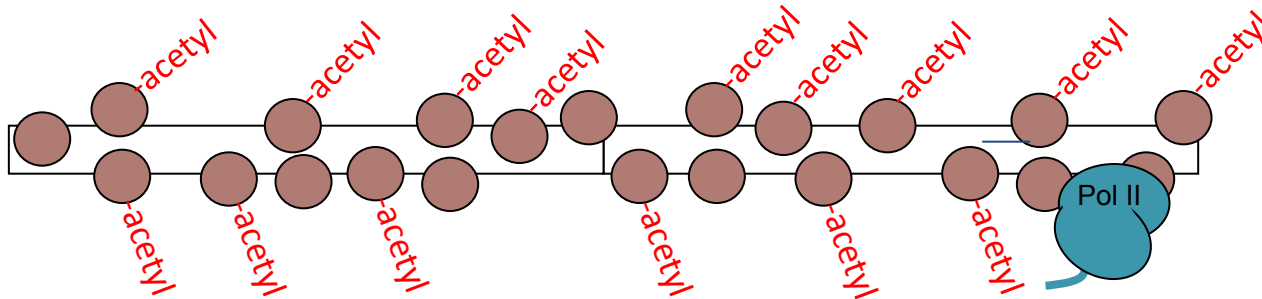
*Chromatin opening should increase SMN2  
E7 inclusion by promoting intragenic Pol II  
elongation: use of histone deacetylase  
inhibitors trichostatin A (TSA) or valproic acid  
(VPA) that, by promoting histone acetylation,  
will open the chromatin*

## COMPACT CHROMATIN (CLOSED)



Valproic acid (VPA)  
"Chromatin opener"

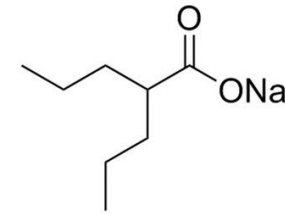
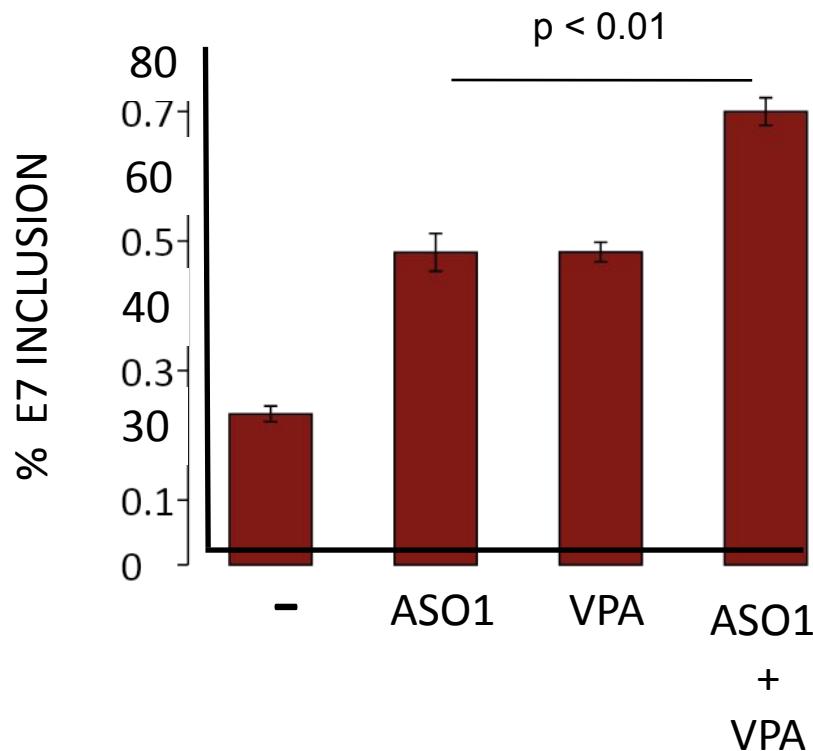
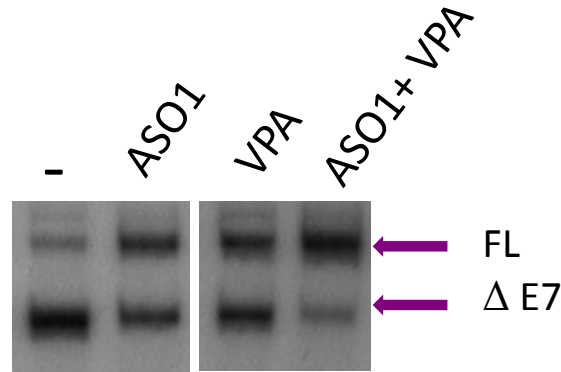
## RELAXED CHROMATIN (OPEN)



# ASO1 (Spinraza) and VPA act cooperatively to promote SMN2 E7 inclusion

HEK293 cells

RT-PCR

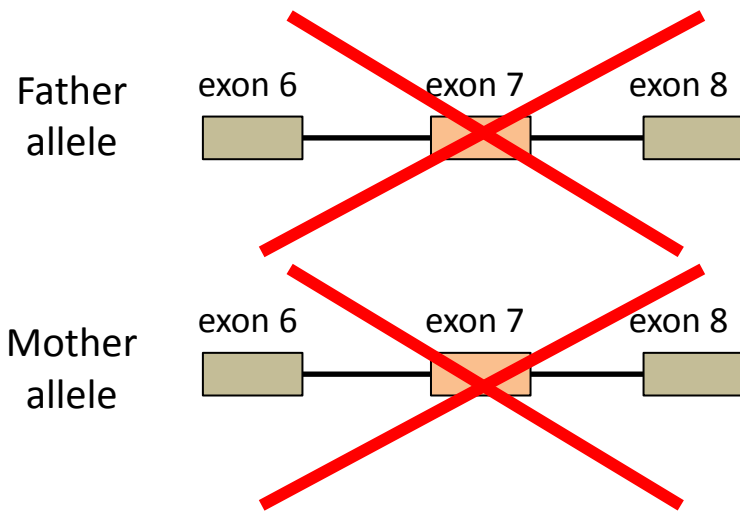


Valproic acid (VPA)  
Class I and II HDACs  
FDA approved  
Similar results with TSA and  
SAHA and in patient  
fibroblasts

*Experiments with an SMA mouse model*

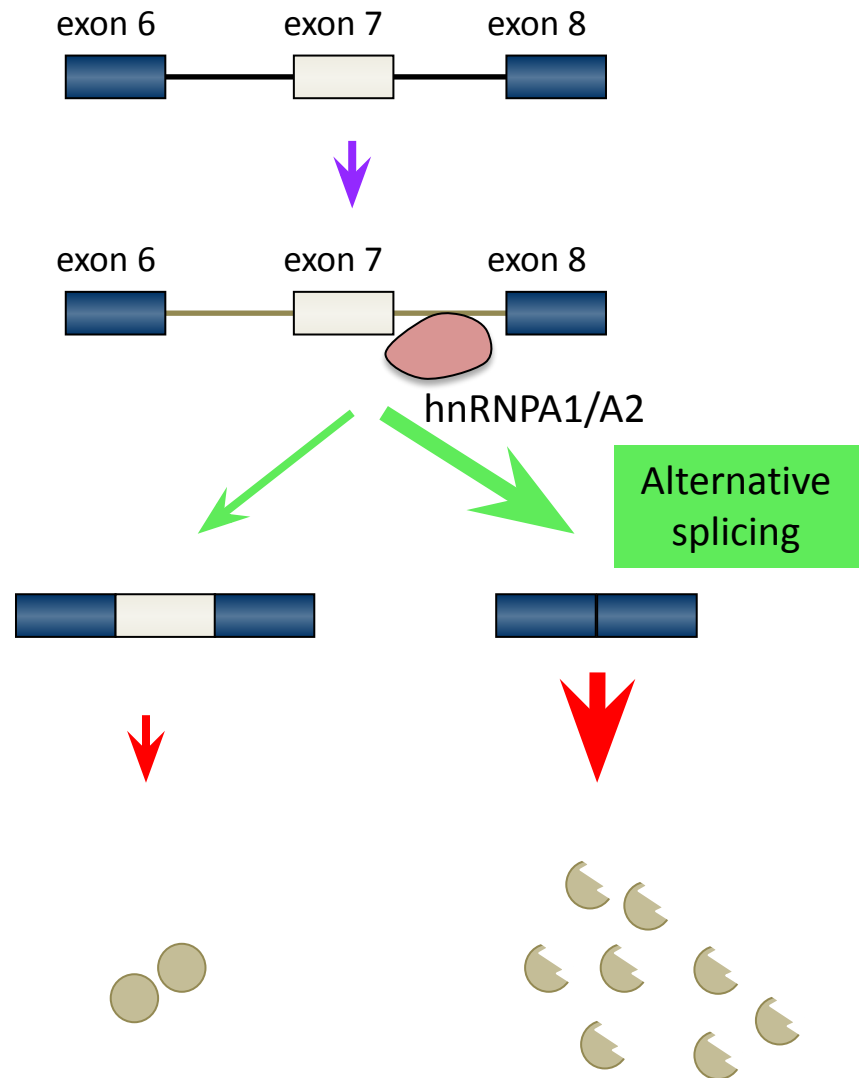
*(Taiwan strain)*

## Only SMN mouse gene



embryonic lethal

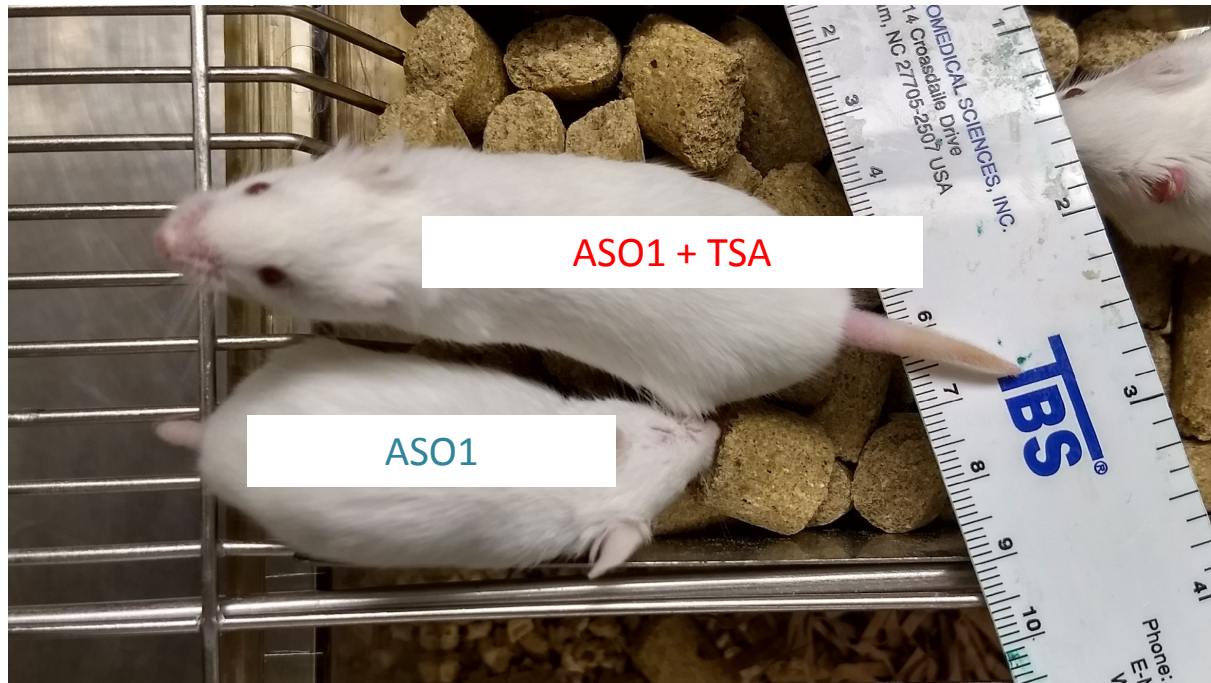
## Human SMN2 transgene



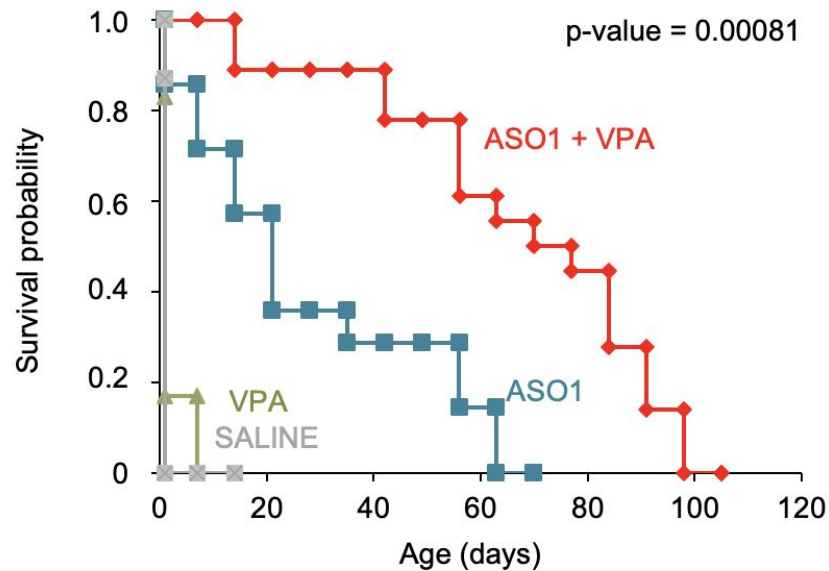
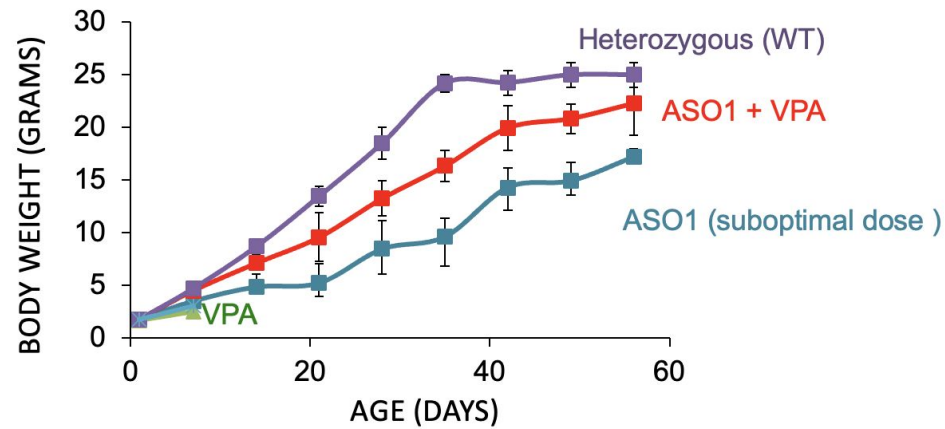
*Single subcutaneous injection of nusinersen-like  
ASO1 (18  $\mu\text{g/g}$ ) and/or HDAC inhibitor (10  $\mu\text{g/g}$ )  
1-2 days after birth*

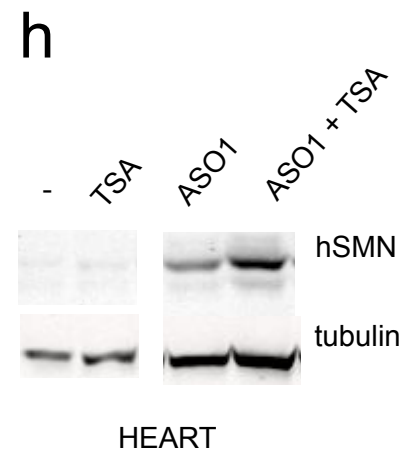
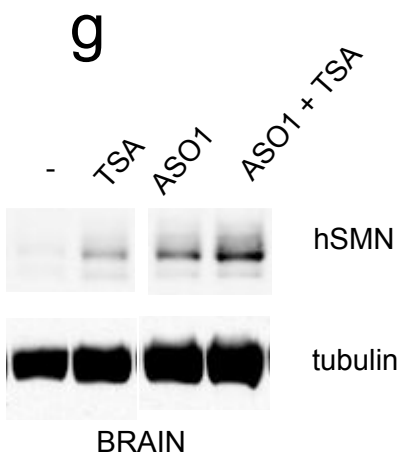
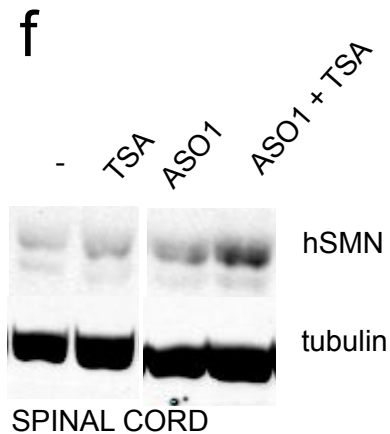
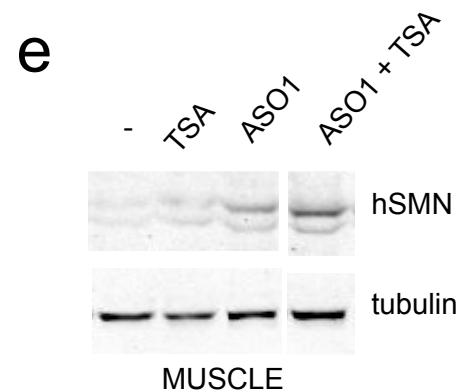
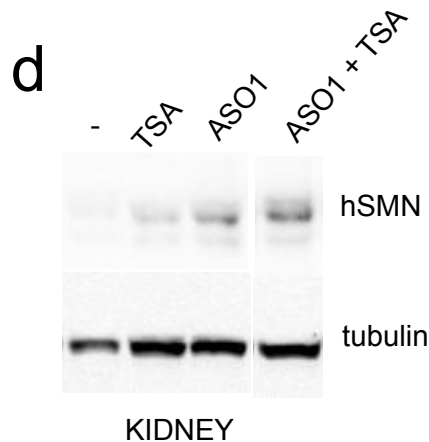
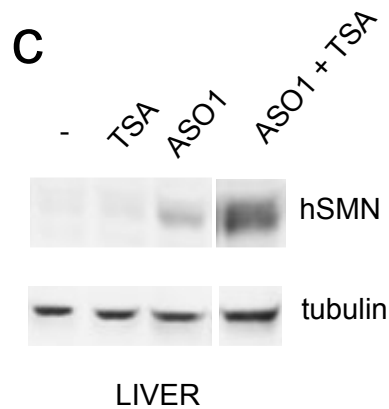
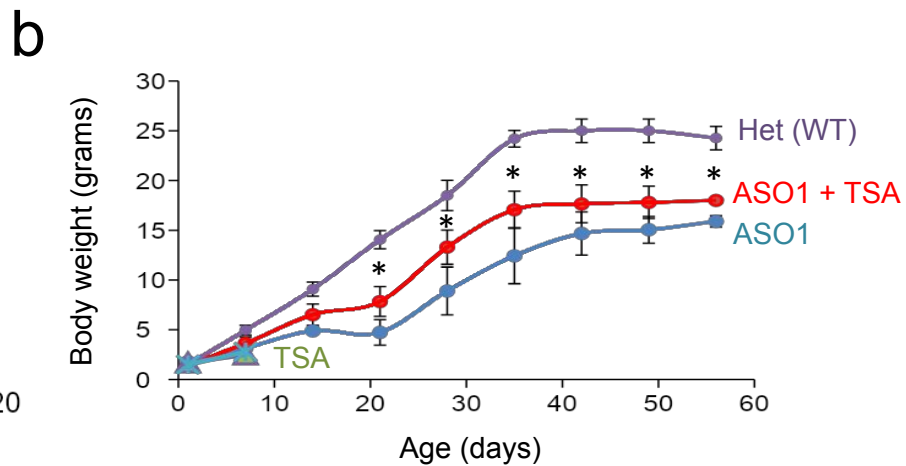
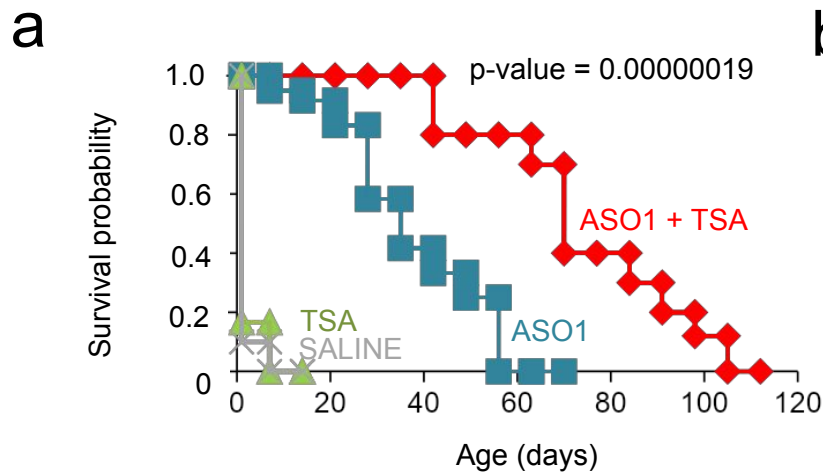
# *Spinraza (ASO1) and HDACs inhibitor act cooperatively in SMA mice model*

42 days old



# *ASO1 and VPA act cooperatively in SMA mice model*





## *Neuromuscular function tests*

# Surface righting

11 days old

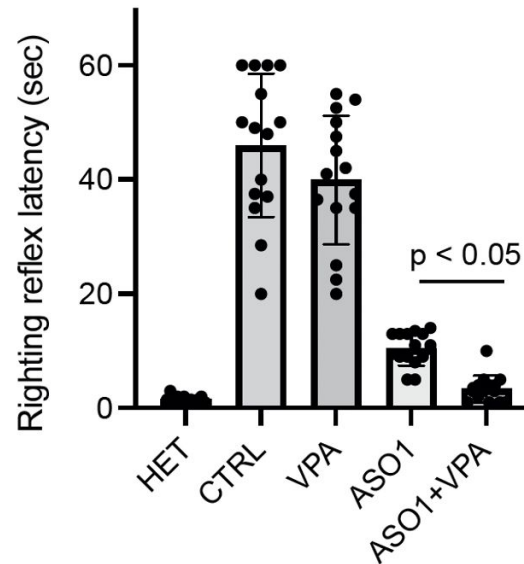
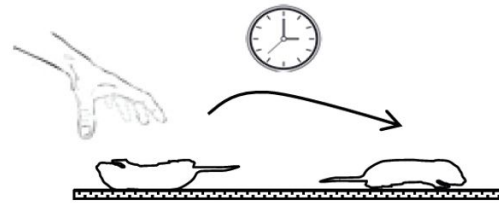


ASO1



ASO1 + VPA

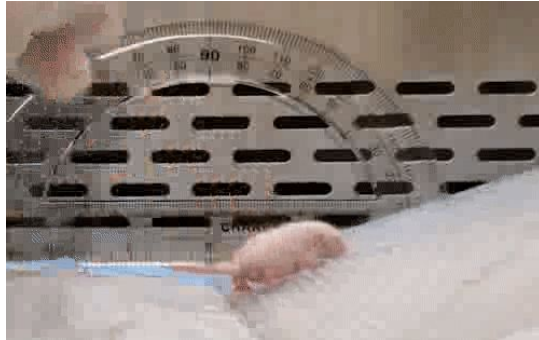
n = 10 (3 tests each)



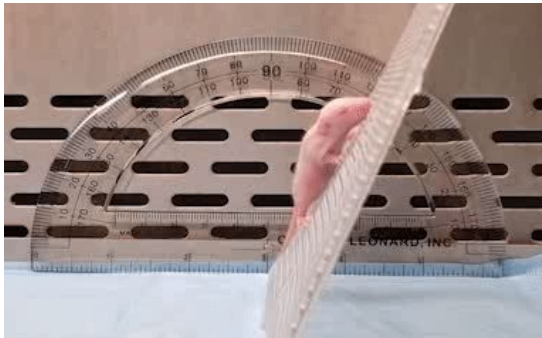
# Grip strength

P7

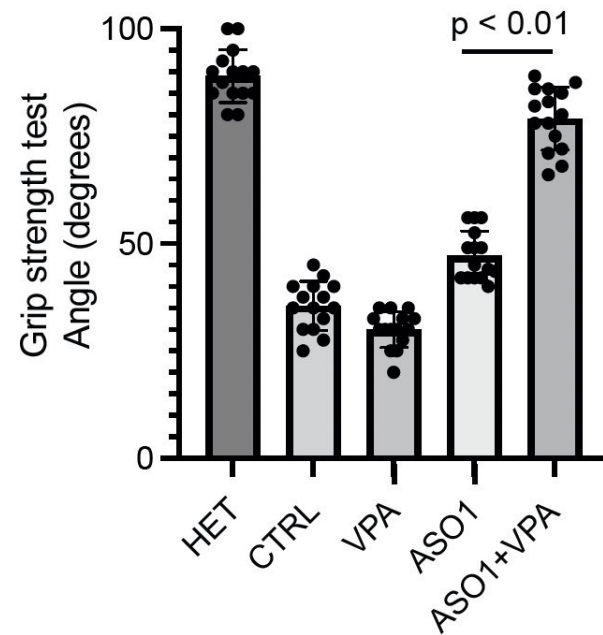
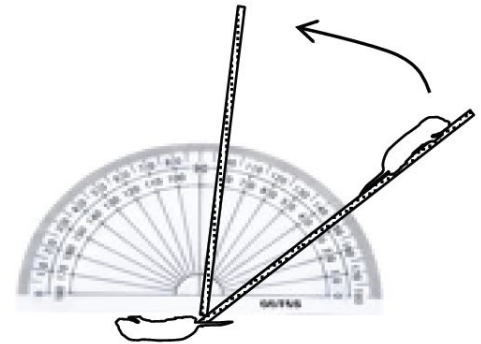
ASO1



ASO1 + VPA



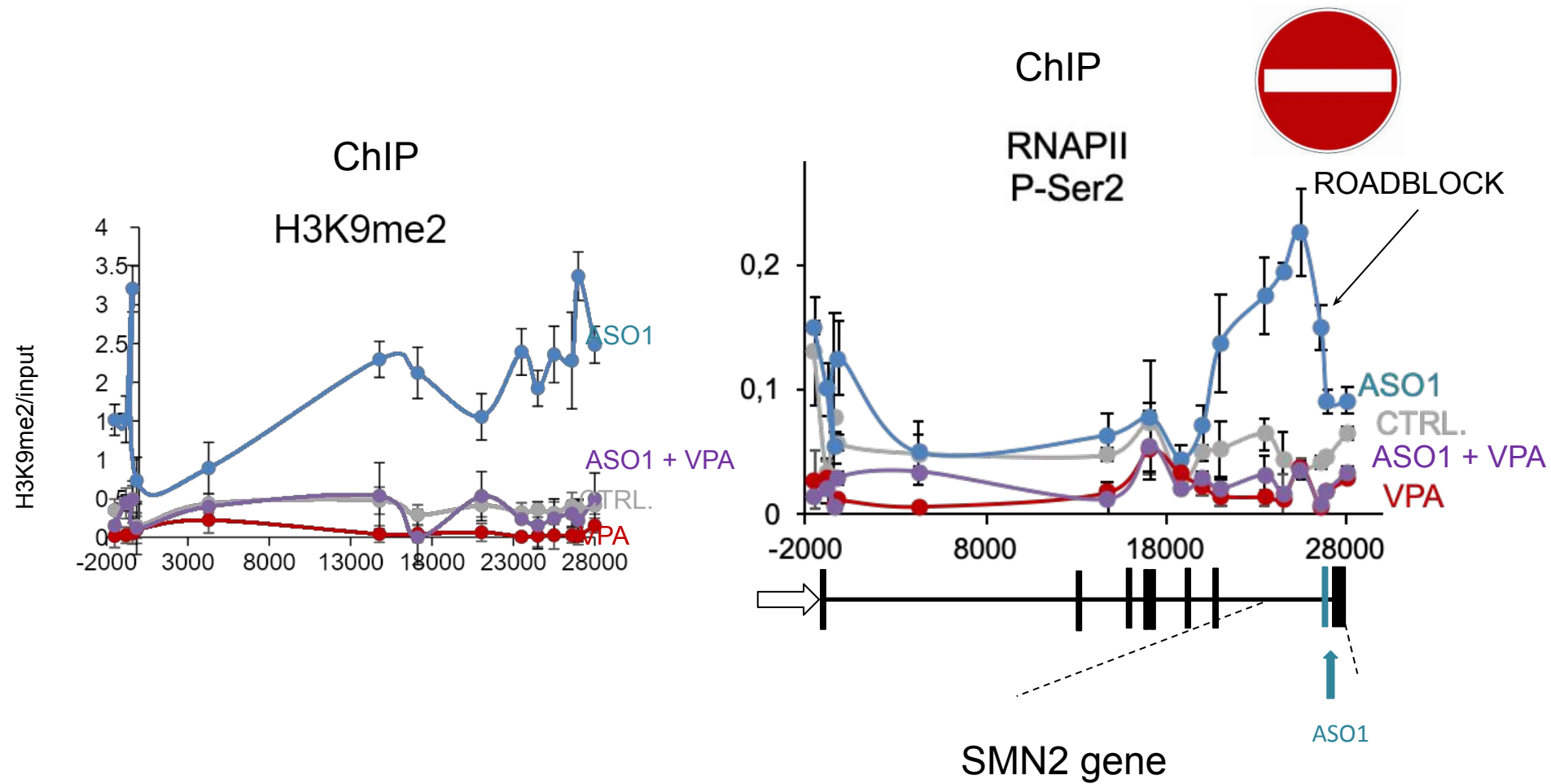
D



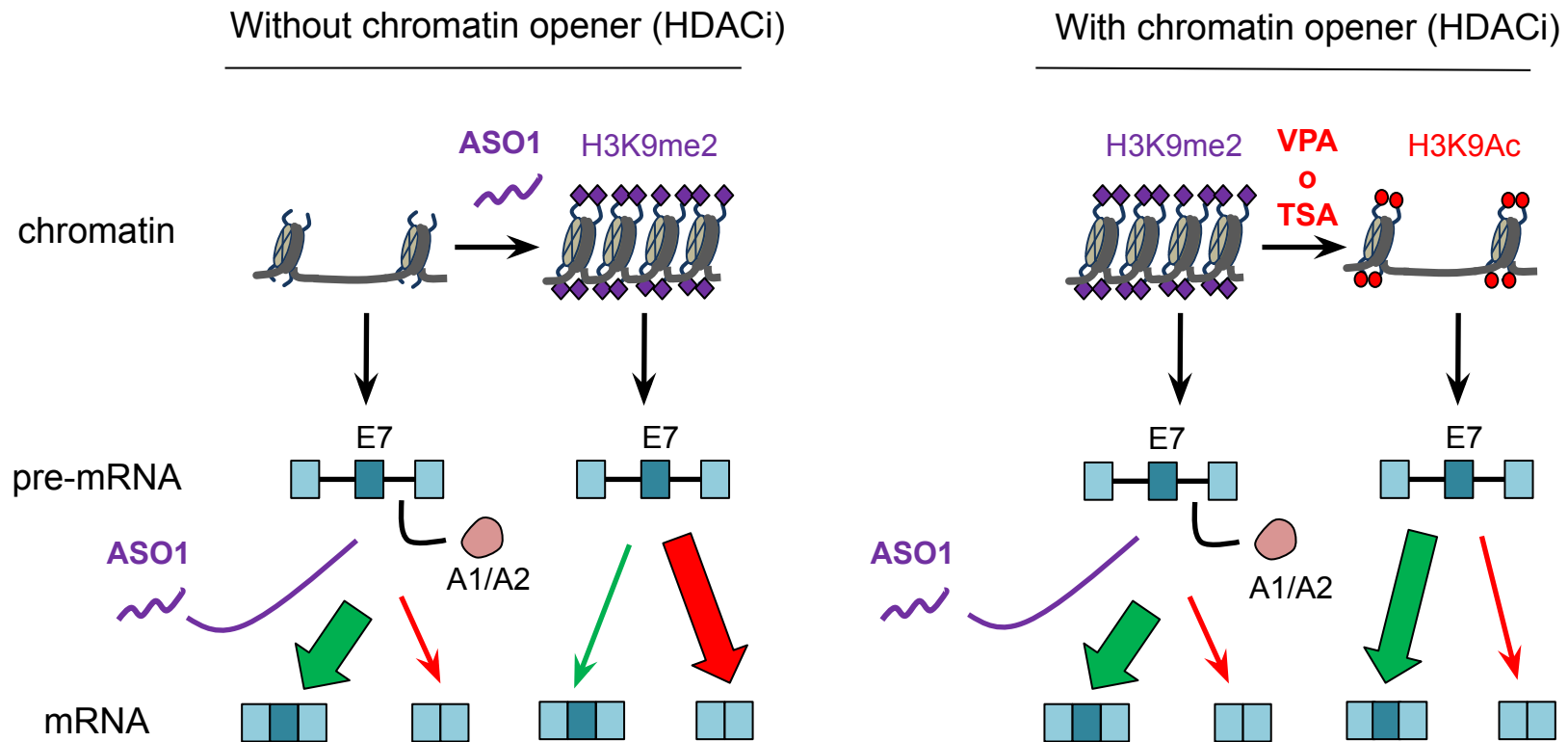
*Chromatin effects of the ASO*

*ASO1 creates a roadblock to  
elongation*

# ASO1 (nusinersen-like) creates a roadblock to PolII elongation



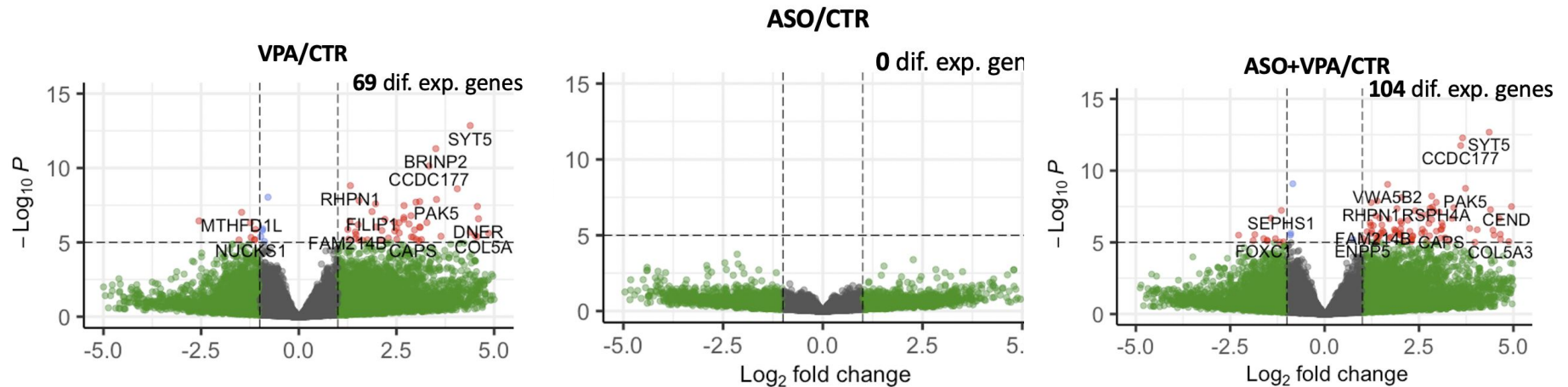
# ASO1 has two opposite effects



*How pleiotropic is VPA?*

# RNA-seq in HEK293 cells

## RNA-seq HEK (gene expression)

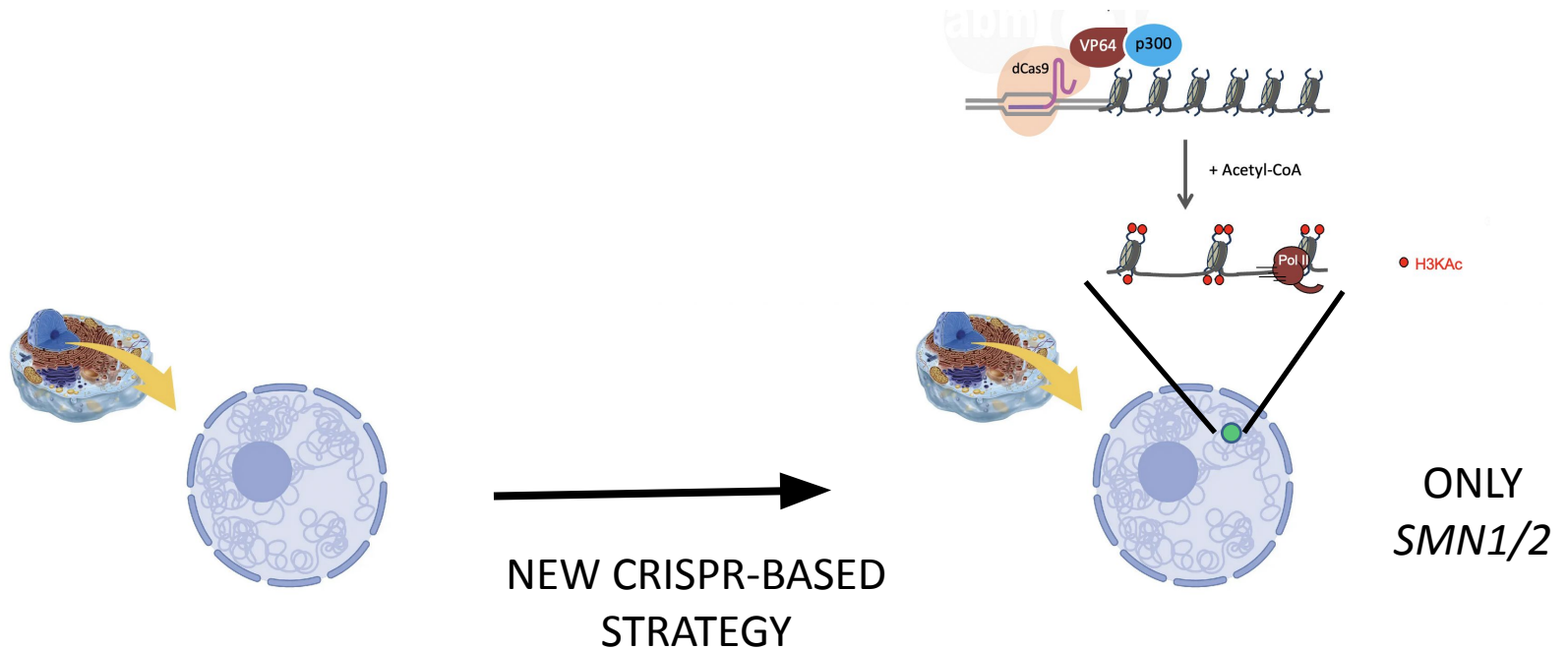
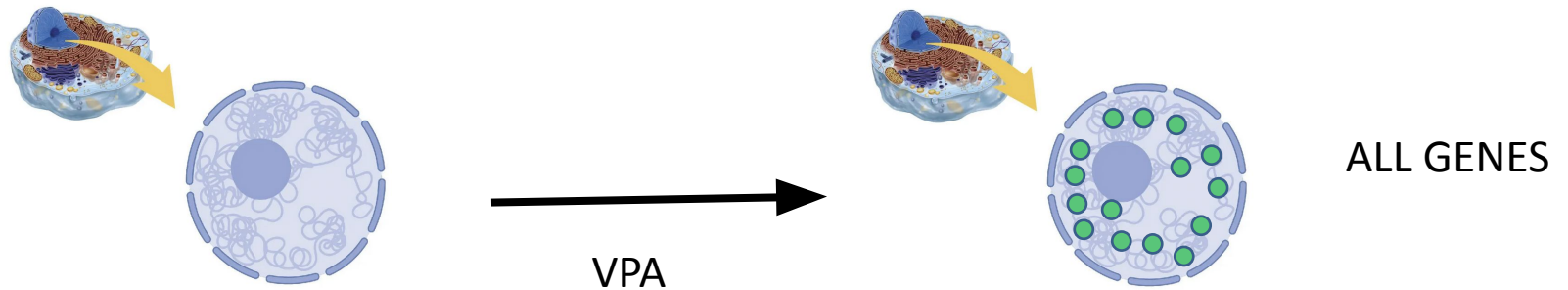


*Anyway and just in case:*

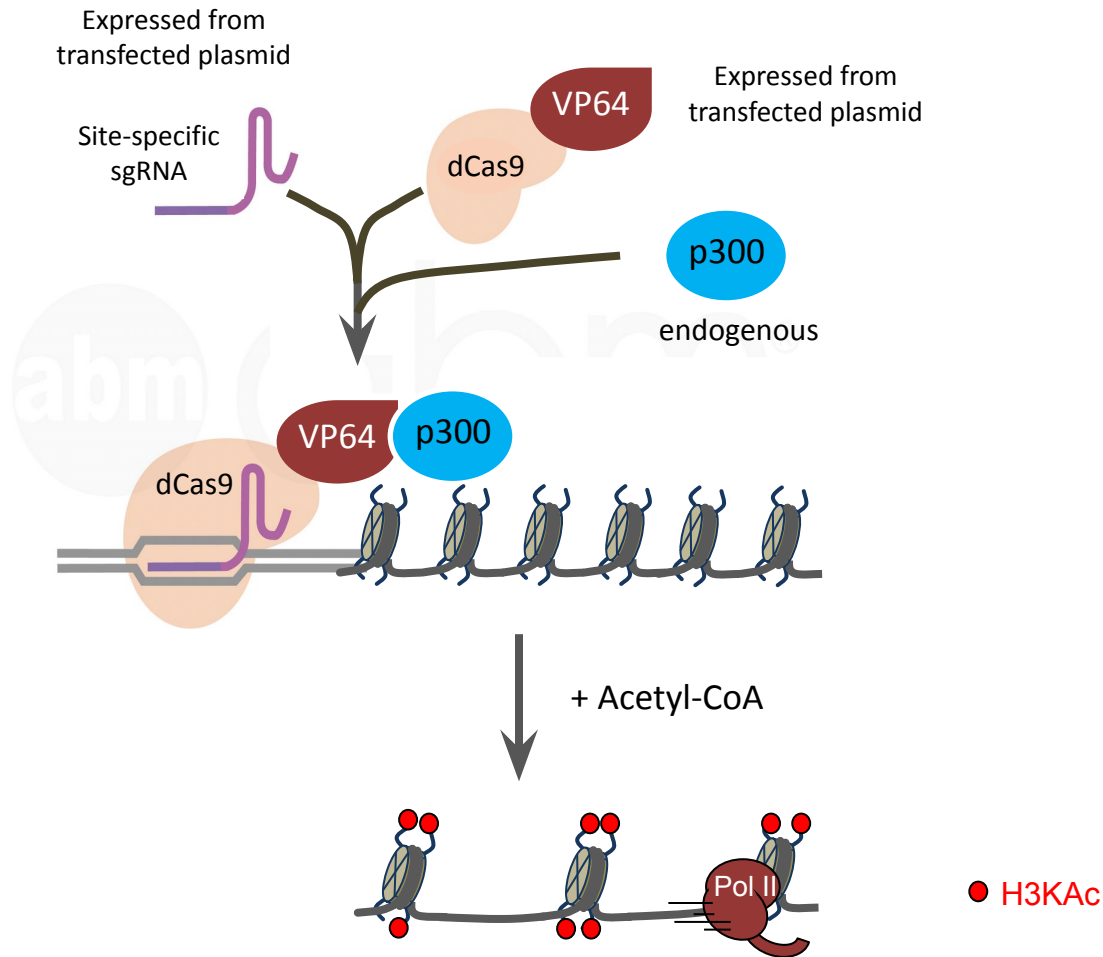
*Can we target H3K9 acetylation just to the SMN2 gene?*

# New strategy

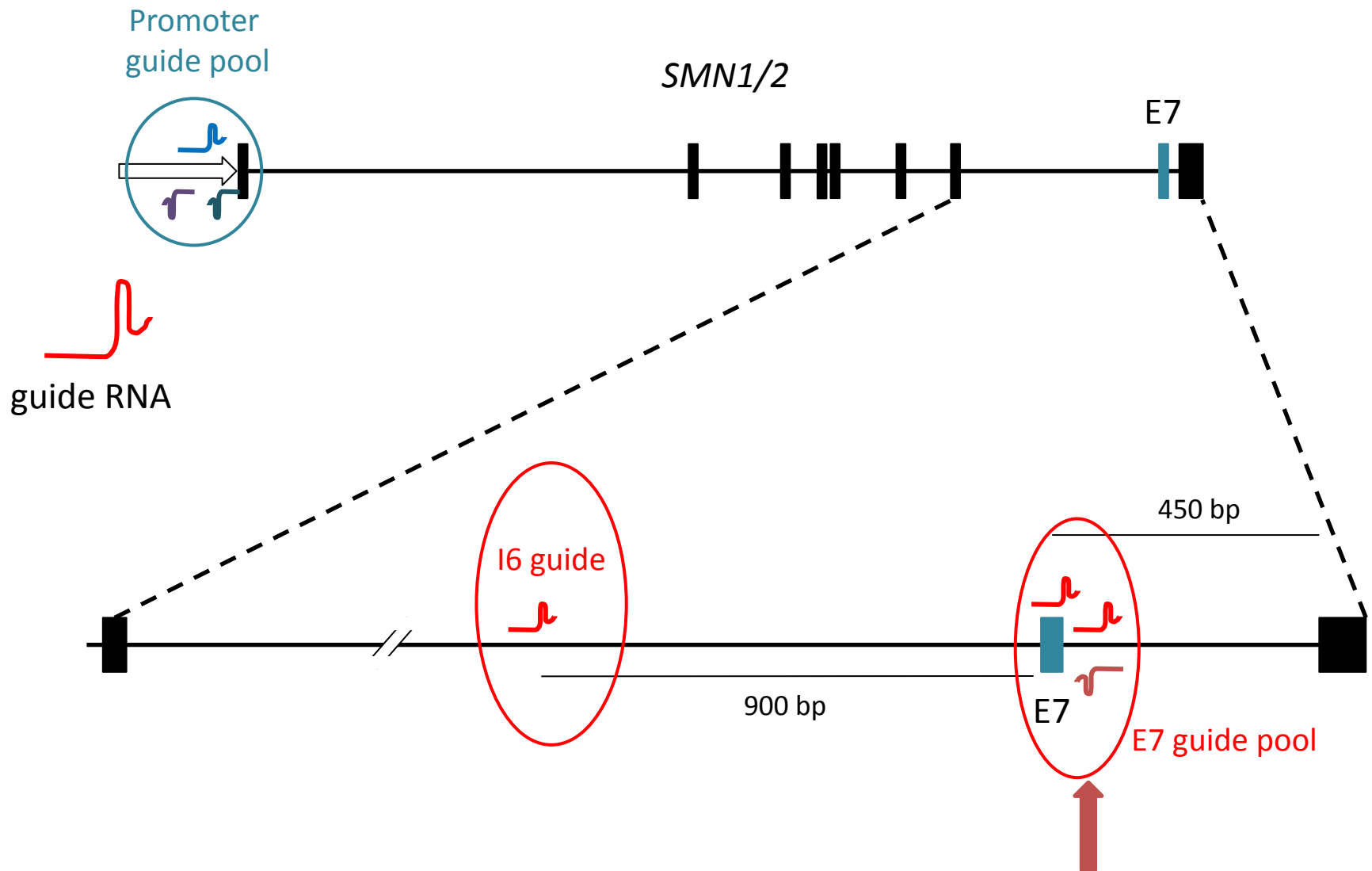
● ACETYLATION = OPEN CHROMATIN



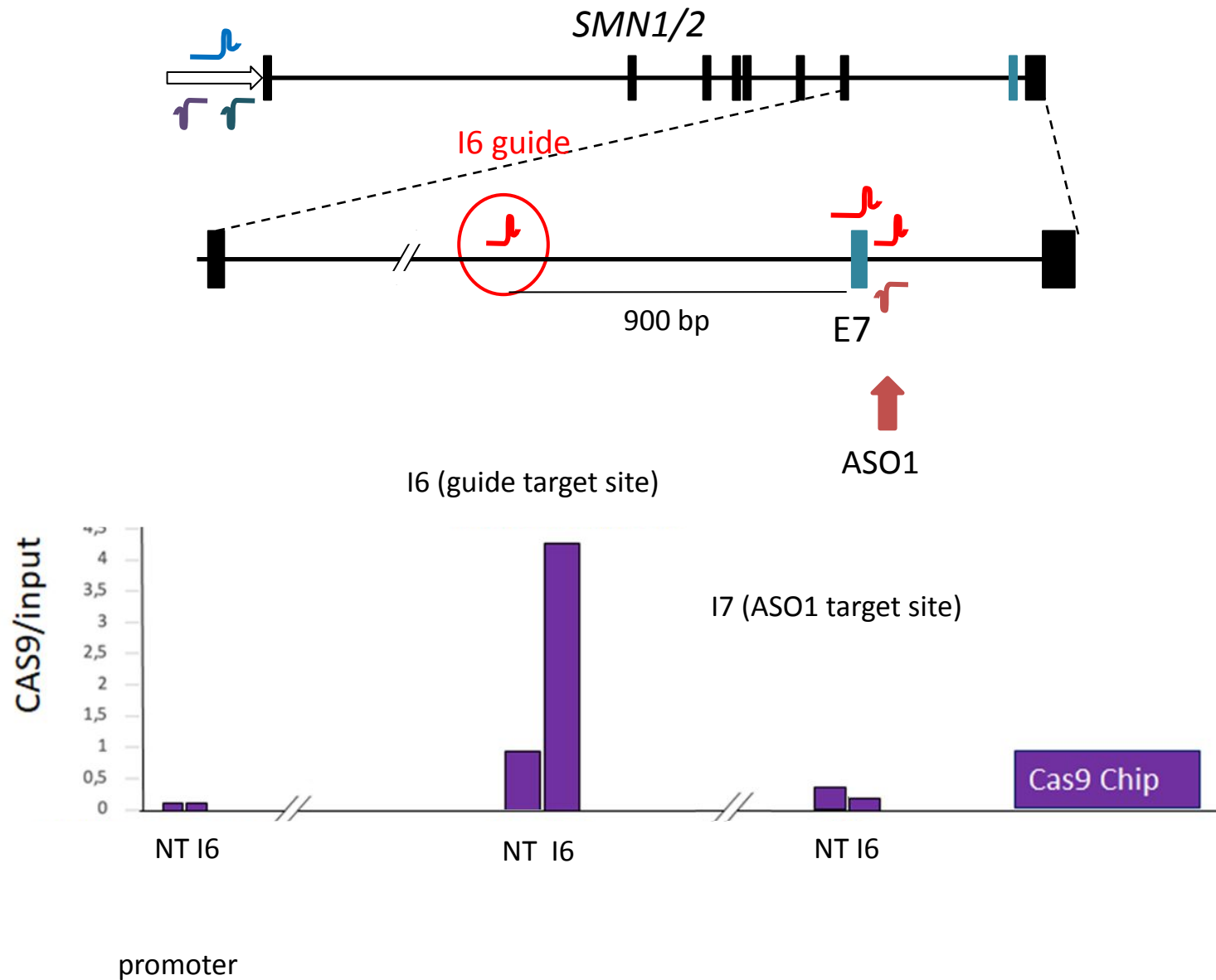
# Dead Cas9 (dCas9) strategy



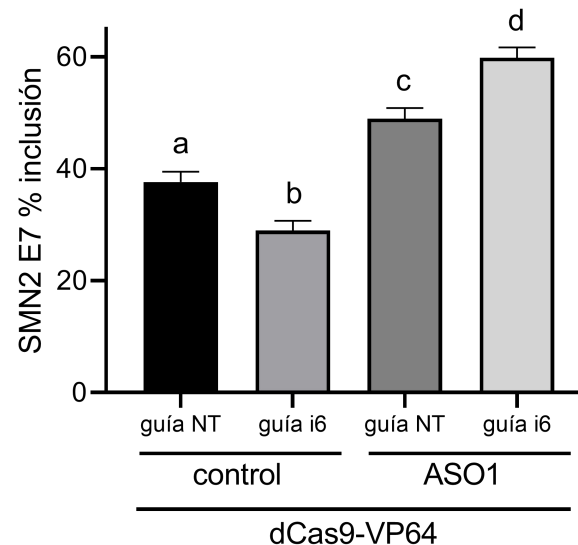
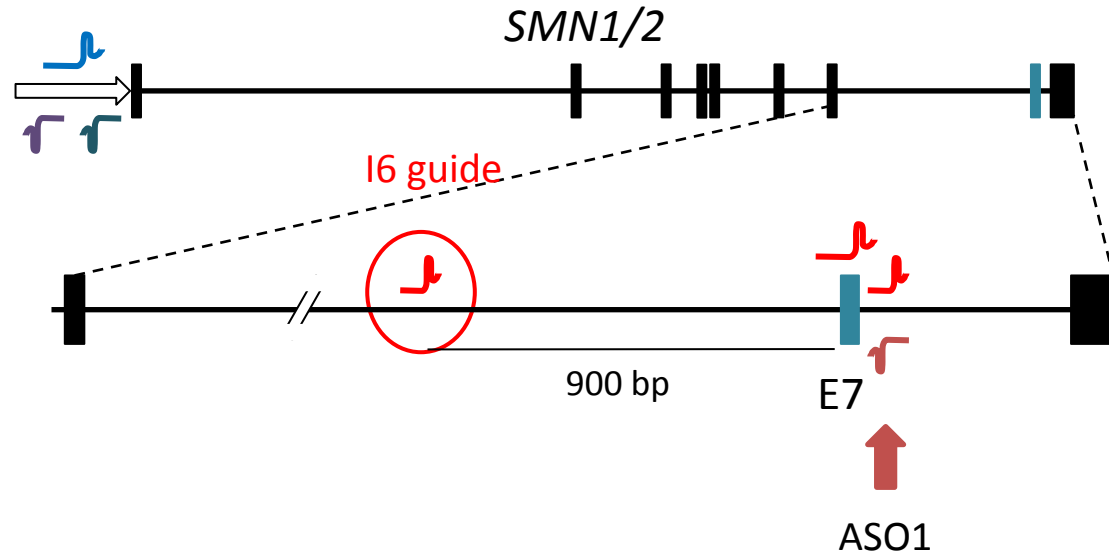
# Dead Cas9 (dCas9) strategy



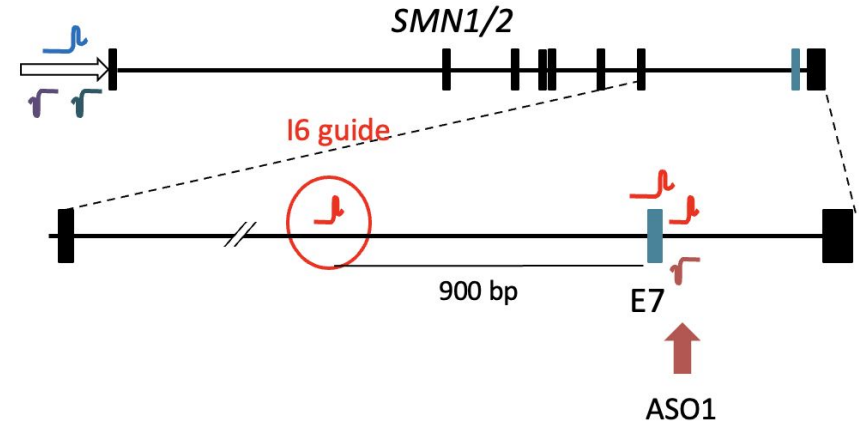
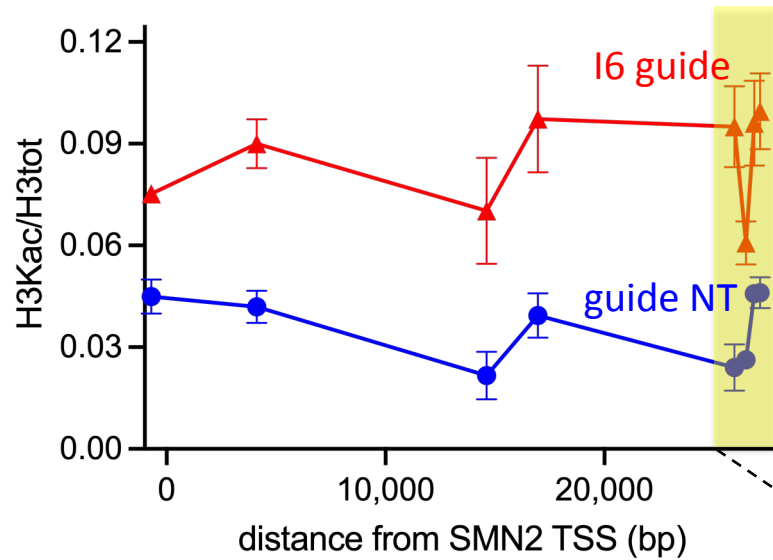
# *dCAs9-VP64 is specifically recruited to the guide target site*



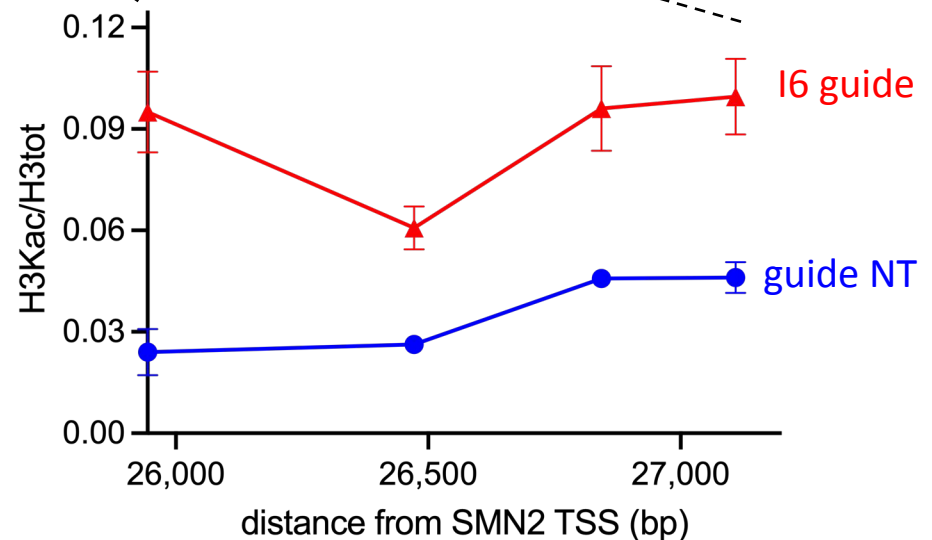
# *dCas9-VP64 with I6 guide cooperate with Spinraza (ASO1)*



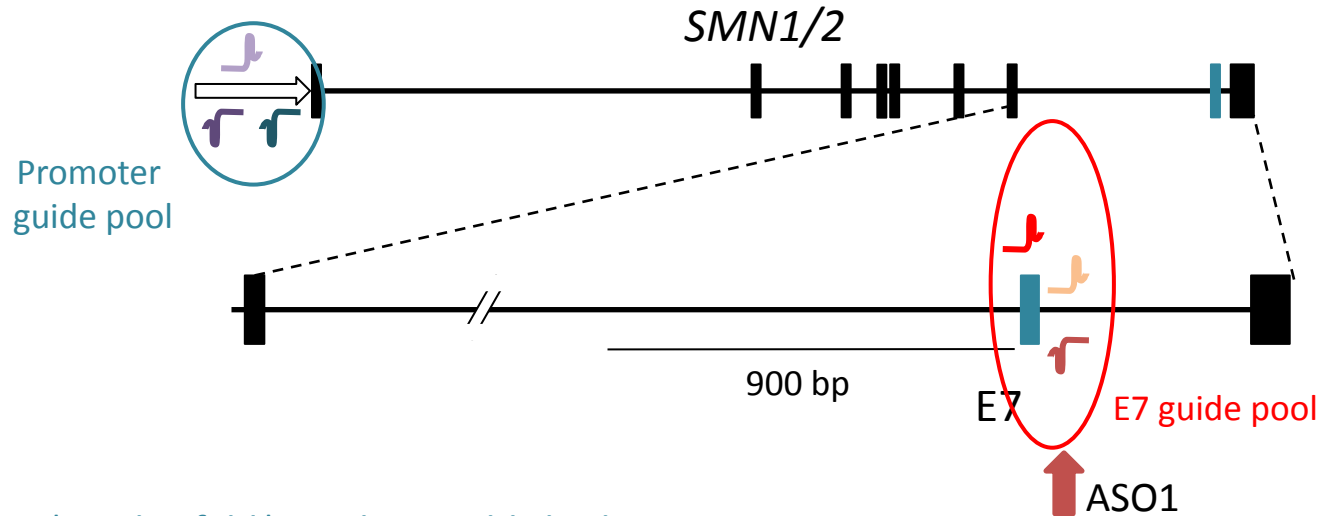
# *Intron 6 guide promotes histone acetylation along the entire SMN1/2 gene*



ChIP  
H3Kac



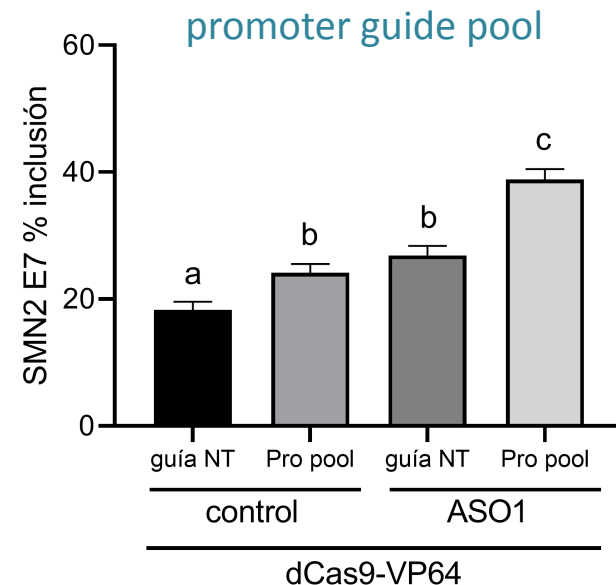
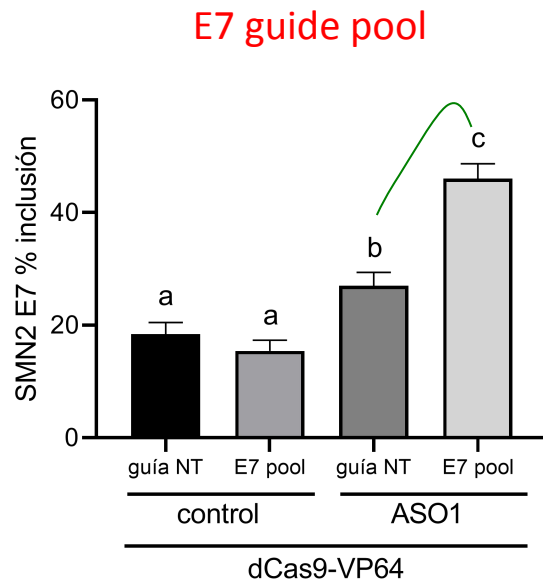
# SMN2-specific acetylation cooperates with Spinraza (ASO1)



Stigliano\*, Haberfeld\* *et al.*, unpublished

## SMN2 E7 % splicing

RT-PCR



## *Conclusions*

*SMN2* E7 is a type II exon: Slow transcript elongation promotes its skipping while fast elongation promotes its inclusion.

Combined therapy for Spinal muscular atrophy: Spinraza (ASO1) and chromatin opening with histone deacetylase inhibitors (VPA).

Mechanism: Spinraza (ASO1) has two opposite effects. The negative effect is counteracted by opening the chromatin with histone deacetylase inhibitors.

Targeting histone acetylation specifically to *SMN1/2* also enhances the effect of Spinraza.



*Designed by Luciana Giono,  
inspired in Alexander Calder  
mobiles*

Marasco, L. E.\*, Dujardin, G.\*, Sousa Luís, R., Hsiu Liu, Y., Stigliano, J., Nomakuchi, T., Proudfoot, N. J., Krainer, A. R. & Kornblihtt, A. R. Counteracting chromatin effects of a splicing-correcting antisense oligonucleotide improves its therapeutic efficacy in spinal muscular atrophy. **Cell** 185, 2057-2070 (2022).



Adrián Krainer

S



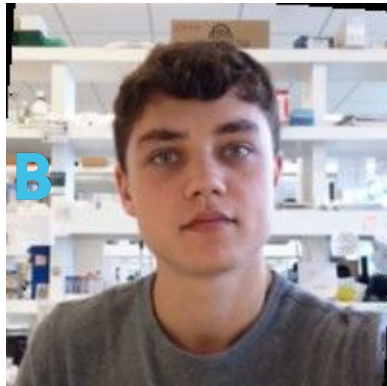
Luciano  
Marasco



Gwendal Dujardin



Nick Proudfoot



Jose  
Stigliano

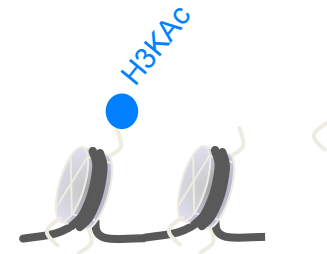
B



Emilia  
Haberfeld



Rui Sousa Luís



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ANPCyT (Argentina)

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CONICET (Argentina)

Lounsbery Foundation (USA)

hhmi

2002-2017

cure  
SMA



RICHARD  
LOUNSBERY  
FOUNDATION





iGracias!

*Chromatin effect of ASOs: DNA or RNA?*

# The chromatin effect seems to be due to interaction of the ASO with DNA

